

# 1 Behaviour Revised: Contaminant Effects on Aquatic Animal Behaviour

2  
3 Greg Pyle<sup>1</sup> and Alex T. Ford<sup>2</sup>

4 | <http://dx.doi.org/10.1016/j.aquatox.2016.11.008>

5  
6 1. Dept. of Biological Sciences, University of Lethbridge, 4401 University Dr.,  
7 Lethbridge, AB, Canada, T1K 3M4

8 2. School of Biological Sciences, University of Portsmouth, King Henry Building,  
9 Portsmouth, UK. PO1 2DT

10  
11 The field of aquatic ecotoxicology has matured over the past several decades. It has  
12 gone from simple “pickle jar” science in the 1960s and ‘70s, to a much more  
13 sophisticated and mechanistic discipline with the advent of molecular and genomic tools.  
14 A long-standing challenge for aquatic ecotoxicology has always been to extrapolate  
15 results generated under controlled laboratory conditions to the variable conditions of the  
16 field. Attempts at making lab-to-field extrapolations have had mixed success. Laboratory  
17 studies conducted at the molecular level have had difficulties translating to natural  
18 conditions, whereas field studies often lack a mechanistic understanding of potential  
19 toxicity.

20 Behavioural toxicology has emerged as a promising discipline to bridge the  
21 laboratory-to-field divide. Animal behaviour integrates the internal physiology of the  
22 animal and the external conditions of the environment. Toxicant-induced behavioural  
23 impairments often point to underlying physiological deficits that can be used effectively  
24 to evaluate ecological risk—especially if the affected behaviour relates directly to  
25 survival, growth, or reproduction.

26 Environmental regulators and industry have been reluctant to adopt behavioural  
27 endpoints for risk assessment purposes, mainly because of a current lack of standardized

28 approaches or perceived difficulties extrapolating lab-based results to natural receiving  
29 environments. However, the ecological relevance of behavioural endpoints is clear given  
30 that they relate directly to survival, growth, and reproduction. In an attempt to advance  
31 the field of behavioural toxicology, particularly as it relates to environmental compliance  
32 and risk assessment, we held a special symposium entitled, “Behaviour Revised:  
33 Contaminant Effects on Aquatic Animal Behaviour,” at the 25th annual meeting of the  
34 Society of Environmental Toxicology and Chemistry (SETAC) Europe meeting, held in  
35 Barcelona, Spain (May 3 - 7, 2015). We invited a range of behavioural toxicologists from  
36 around the world to contribute towards advancing our understanding of how  
37 environmental toxicants affect animal behaviours with a mind towards advancing the  
38 discipline for the purposes of environmental regulation and ecological risk assessment. It  
39 was at this meeting that we began soliciting manuscripts for this special issue of *Aquatic*  
40 *Toxicology*, both from the participants at the special symposium, as well as from other  
41 experts from around the world. The collection of papers in this special issue is from those  
42 that answered the call.

43       Increased pharmaceutical use in humans and domestic animals has led to a  
44 concomitant increase in the presence of the same pharmaceuticals and their metabolites in  
45 aquatic systems. Pharmaceuticals are now considered to be ‘emerging’ contaminants of  
46 concern in aquatic ecosystems. Henriques et al. (2016) report that locomotor function is  
47 impaired in zebrafish embryos exposed to gemfibrozil—a pharmaceutical lipid  
48 regulator—at concentrations that are significantly lower than those required to induce  
49 developmental anomalies. Similarly low concentrations of carbamazepine, diazepam,  
50 propranolol, and fluoxetine, in the ng/L range, were sufficient to impair the phototactic

51 response of *Daphnia magna* (Rivetti et al., 2016). Androgenic growth hormones, such as  
52 17 $\beta$ -trenbolone are frequently administered to cattle as growth promoters. However, as  
53 Tomkins et al. (2016) demonstrate, concentrations as low as 4 ng/L are sufficient to affect  
54 female mate choice in wild guppies. These pharmaceutical-induced effects on locomotion  
55 and phototaxis could result in an impaired ability to respond to predators. Effects on  
56 female mate choice could affect reproductive output as well as the genetic integrity of  
57 offspring.

58 Metals are also well known neurotoxicants that can affect animal behaviour. Pereira  
59 et al. (2016) linked changes in white sea bream brain morphology following exposure to  
60 inorganic mercury to impaired motor function and anxiety status. Although some brain  
61 regions could recover once the mercury was removed, others could not, resulting in some  
62 long-term behavioural deficits. Ašmonaitė et al. (2016) present a novel approach using  
63 early life-stage zebrafish locomotion from video-tracking software and multivariate  
64 statistical analyses to develop heat maps for fish exposed to both silver ions and  
65 nanoparticles. Their approach allowed them to differentiate ion effects from  
66 nanoparticles, and different behavioural profiles associated with exposure concentrations;  
67 hypoactivity at high concentrations, and hyperactivity at low concentrations. Gauthier et  
68 al. (2016) followed-up a previous study (Gauthier et al., 2015) that showed that mixtures  
69 of copper and phenanthrene (a polycyclic aromatic hydrocarbon; PAH) causes  
70 potentiated toxicity in the freshwater scud, *Hyalella azteca*. Here, they demonstrate that  
71 copper and phenanthrene act independently to induce toxicity, and the behavioural  
72 deficits induced by phenanthrene exposure (e.g., hyperactive, uncoordinated activity)  
73 result from an acetylcholinesterase inhibition, much like an organophosphate pesticide.

74 Pesticides can also lead to several behavioural toxicological effects. Andrade et al.  
75 (2016) exposed early life-stage zebrafish to carbendazim, a broad-spectrum fungicide,  
76 and demonstrated that behavioural impairments occurred at exposure concentrations that  
77 were orders of magnitude lower than those required to affect development, metabolism,  
78 or lethality. Rodrigues et al. (2016) exposed freshwater planaria to chlorantraniliprole—  
79 an anthranilic diamide insecticide—and demonstrated impaired feeding and swimming  
80 behaviours using a video tracking system at low, ecologically relevant exposure  
81 concentrations. Renick et al. (2016) tested the interactive effects of an organophosphate  
82 pesticide, chlorpyrifos, and a common trematode parasite, *Euhaplorchis californiensis*, in  
83 Californian coastal waters on the California killifish. The pesticide reduced the activity of  
84 the neurotransmitter acetylcholinesterase, resulting in reduced swimming speeds; whereas  
85 the trematode had no effect on swimming behaviour.

86 Contaminants associated with municipal and industrial waste waters can also cause  
87 behavioural effects in aquatic organisms. Lanctôt et al. (2016) demonstrated that striped  
88 marsh frog tadpoles exposed to coal mine waste water for a short duration (24 h) became  
89 hyperactive. However, this effect was transient and was not detectable after 28 days of  
90 exposure. Lari et al. (2016) showed that Caspian roach exposed to the water soluble  
91 fraction of crude oil show impaired swimming performance, which could be linked to  
92 contaminant-induced impairments in the oxygen delivery system (e.g., decreased pO<sub>2</sub> and  
93 increased pCO<sub>2</sub> in oil exposed animals). Melvin (2016) showed increased swimming  
94 velocity and body condition, and decreased lipid content in juvenile empire gudgeons  
95 exposed to full-strength municipal waste water.

96 The recent emergence of behavioural toxicology as a viable approach to integrate lab-

97 and-field has given rise to a number of different tools that will be useful as we begin to  
98 develop into the area of regulatory compliance. Hartmann et al. (2016) present a novel  
99 method for using freshwater mussel behaviour, such as filtration, valve transition  
100 frequency (from open to closed), and contaminant avoidance, as viable ecotoxicological  
101 endpoints for chemical exposures, temperature changes, and particulate matter  
102 contamination. Kristofco et al. (2016) address the lack of high-throughput screening  
103 assays available for behavioural toxicology, and demonstrate that understanding specific  
104 life-stage responses can lead to more sensitive behavioural toxicological endpoints. By  
105 testing several zebrafish developmental stages, they demonstrate that larvae 10 days post  
106 hatch (dph) are significantly more sensitive to diazinon (pesticide) and diphenhydramine  
107 (pharmaceutical) than earlier, embryonic life-stages known to be more sensitive in  
108 growth and survival tests. Kaluef et al. (2016) argue that zebrafish neurobehavioural  
109 phenomics is a promising new approach that integrates behavioural phenotypes with  
110 various genetic and environmental factors. They specifically advocate for zebrafish as a  
111 model owing to its homology with the human genome and the wide range of genomic and  
112 molecular tools available for immediate experimentation. Michalec et al. (2016) describe  
113 the use of lipid nanocapsules to deliver hydrophobic toxicants (PCBs and PAHs) to  
114 calanoid copepods. The new delivery system allows for an analysis of the relative  
115 contribution of water and dietary toxicant exposure routes as they might affect 3-D  
116 copepod swimming behaviour.

117 Several authors in this issue provide critical reviews related to different aspects of  
118 behavioural toxicology. Parker (2016) argues against methodological standardization for  
119 behavioural toxicology assays given the numerous confounding variables that can

120 influence animal behaviour. Instead, he offers approaches that make use of the test  
121 animal's unconditioned natural behavioural tendencies, which can increase the reliability  
122 and ecological validity of behavioural analyses. Hellström et al. (2016) provide a review  
123 of current acoustic telemetry tools available to monitor animal behaviour in natural  
124 systems. These tools provide a direct link between results generated under tightly  
125 controlled laboratory conditions to the realities of natural systems. Tierney (2016)  
126 reviews the literature on fish behavioural avoidance responses to contaminants in the  
127 environment. He demonstrates that avoidance responses are common, but not universal.  
128 Several studies have documented cases where fishes are attracted to contaminants at  
129 concentrations that are likely maladaptive. He then provides a framework on which to  
130 base future studies to screen fish species for their ability to detect and avoid  
131 environmental contaminants.

132       Together, these papers comprise a snapshot of the current state of aquatic behavioural  
133 toxicology. Advances in our ability to track animal behaviour using sophisticated video  
134 and high-throughput tracking systems has increased the resolution with which  
135 contaminant-induced behavioural deficits can be measured. New animal models and  
136 novel contaminant-delivery systems provide researchers with a well-supplied tool chest  
137 for improving the ecological relevance of environmental risk assessments. Genomic and  
138 molecular tools provide a mechanistic underpinning of behavioural effects resulting from  
139 contaminant exposures. Behavioural toxicology is no longer an emerging field. We now  
140 understand that environmental contaminants—including pharmaceuticals, metals,  
141 pesticides, and municipal and industrial waste waters, among others—can affect aquatic  
142 animal behaviour resulting in maladaptive effects that need to be considered by

143 environmental managers and government regulators. Our next challenges include linking  
144 behavioural effects observed under controlled laboratory conditions to natural systems,  
145 and to adopt well-established contaminant-induced behavioural deficits into regulatory  
146 compliance legislation.

#### 147 **Literature Cited**

- 148 Andrade, T.S., Henriques, J.F., Almeida, A.R., Machado, A.L., Koba, O., Giang, P.T.,  
149 Soares, A.M.V.M., Domingues, I., 2016. Carbendazim exposure induces  
150 developmental, biochemical and behavioural disturbance in zebrafish embryos.  
151 *Aquat. Toxicol.* 170, 390–399. doi:10.1016/j.aquatox.2015.11.017
- 152 Ašmonaitė, G., Boyer, S., Souza, K.B. de, Wassmur, B., Sturve, J., 2016. Behavioural  
153 toxicity assessment of silver ions and nanoparticles on zebrafish using a  
154 locomotion profiling approach. *Aquat. Toxicol.* 173, 143–153.  
155 doi:10.1016/j.aquatox.2016.01.013
- 156 Gauthier, P.T., Norwood, W.P., Prepas, E.E., Pyle, G.G., 2016. Behavioural alterations  
157 from exposure to Cu, phenanthrene, and Cu-phenanthrene mixtures: linking  
158 behaviour to acute toxic mechanisms in the aquatic amphipod, *Hyalella azteca*.  
159 *Aquat. Toxicol.* 170, 377–383. doi:10.1016/j.aquatox.2015.10.019
- 160 Gauthier, P.T., Norwood, W.P., Prepas, E.E., Pyle, G.G., 2015. Metal–polycyclic  
161 aromatic hydrocarbon mixture toxicity in *Hyalella azteca*. 2. Metal accumulation  
162 and oxidative stress as interactive co-toxic mechanisms. *Environ. Sci. Technol.*  
163 49. doi:10.1021/acs.est.5b03233

164 Hartmann, J.T., Beggel, S., Auerswald, K., Stoeckle, B.C., Geist, J., 2016. Establishing  
165 mussel behavior as a biomarker in ecotoxicology. *Aquat. Toxicol.* 170, 279–288.  
166 doi:10.1016/j.aquatox.2015.06.014

167 Hellström, G., Klaminder, J., Jonsson, M., Fick, J., Brodin, T., 2016. Upscaling  
168 behavioural studies to the field using acoustic telemetry. *Aquat. Toxicol.* 170,  
169 384–389. doi:10.1016/j.aquatox.2015.11.005

170 Henriques, J.F., Almeida, A.R., Andrade, T., Koba, O., Golovko, O., Soares, A.M.V.M.,  
171 Oliveira, M., Domingues, I., 2016. Effects of the lipid regulator drug gemfibrozil:  
172 A toxicological and behavioral perspective. *Aquat. Toxicol.* 170, 355–364.  
173 doi:10.1016/j.aquatox.2015.09.017

174 Kalueff, A.V., Echevarria, D.J., Homechaudhuri, S., Stewart, A.M., Collier, A.D.,  
175 Kaluyeva, A.A., Li, S., Liu, Y., Chen, P., Wang, J., Yang, L., Mitra, A., Pal, S.,  
176 Chaudhuri, A., Roy, A., Biswas, M., Roy, D., Podder, A., Poudel, M.K., Katare,  
177 D.P., Mani, R.J., Kyzar, E.J., Gaikwad, S., Nguyen, M., Song, C., 2016. Zebrafish  
178 neurobehavioral phenomics for aquatic neuropharmacology and toxicology  
179 research. *Aquat. Toxicol.* 170, 297–309. doi:10.1016/j.aquatox.2015.08.007

180 Kristofco, L.A., Cruz, L.C., Haddad, S.P., Behra, M.L., Chambliss, C.K., Brooks, B.W.,  
181 2016. Age matters: Developmental stage of *Danio rerio* larvae influences  
182 photomotor response thresholds to diazinion or diphenhydramine. *Aquat. Toxicol.*  
183 170, 344–354. doi:10.1016/j.aquatox.2015.09.011

184 Lanctôt, C., Bennett, W., Wilson, S., Fabbro, L., Leusch, F.D.L., Melvin, S.D., 2016.  
185 Behaviour, development and metal accumulation in striped marsh frog tadpoles

186 (*Limnodynastes peronii*) exposed to coal mine wastewater. *Aquat. Toxicol.* 173,  
187 218–227. doi:10.1016/j.aquatox.2016.01.014

188 Lari, E., Abtahi, B., Hashtroudi, M.S., 2016. The effect of the water soluble fraction of  
189 crude oil on survival, physiology and behaviour of Caspian roach, *Rutilus*  
190 *caspicus* (Yakovlev, 1870). *Aquat. Toxicol.* 170, 330–334.  
191 doi:10.1016/j.aquatox.2015.09.003

192 Melvin, S.D., 2016. Short-term exposure to municipal wastewater influences energy,  
193 growth, and swimming performance in juvenile Empire Gudgeons (*Hypseleotris*  
194 *compressa*). *Aquat. Toxicol.* 170, 271–278. doi:10.1016/j.aquatox.2015.06.003

195 Michalec, F.-G., Holzner, M., Souissi, A., Stancheva, S., Barras, A., Boukherroub, R.,  
196 Souissi, S., 2016. Lipid nanocapsules for behavioural testing in aquatic  
197 toxicology: Time–response of *Eurytemora affinis* to environmental concentrations  
198 of PAHs and PCB. *Aquat. Toxicol.* 170, 310–322.  
199 doi:10.1016/j.aquatox.2015.08.010

200 Parker, M.O., 2016. Adult vertebrate behavioural aquatic toxicology: Reliability and  
201 validity. *Aquat. Toxicol.* 170, 323–329. doi:10.1016/j.aquatox.2015.09.001

202 Pereira, P., Puga, S., Cardoso, V., Pinto-Ribeiro, F., Raimundo, J., Barata, M., Pousão-  
203 Ferreira, P., Pacheco, M., Almeida, A., 2016. Inorganic mercury accumulation in  
204 brain following waterborne exposure elicits a deficit on the number of brain cells  
205 and impairs swimming behavior in fish (white seabream—*Diplodus sargus*).  
206 *Aquat. Toxicol.* 170, 400–412. doi:10.1016/j.aquatox.2015.11.031

207 Renick, V.C., Weinersmith, K., Vidal-Dorsch, D.E., Anderson, T.W., 2016. Effects of a  
208 pesticide and a parasite on neurological, endocrine, and behavioral responses of

209 an estuarine fish. *Aquat. Toxicol.* 170, 335–343.  
210 doi:10.1016/j.aquatox.2015.09.010

211 Rivetti, C., Campos, B., Barata, C., 2016. Low environmental levels of neuro-active  
212 pharmaceuticals alter phototactic behaviour and reproduction in *Daphnia magna*.  
213 *Aquat. Toxicol.* 170, 289–296. doi:10.1016/j.aquatox.2015.07.019

214 Rodrigues, A.C.M., Henriques, J.F., Domingues, I., Golovko, O., Žlábek, V., Barata, C.,  
215 Soares, A.M.V.M., Pestana, J.L.T., 2016. Behavioural responses of freshwater  
216 planarians after short-term exposure to the insecticide chlorantraniliprole. *Aquat.*  
217 *Toxicol.* 170, 371–376. doi:10.1016/j.aquatox.2015.10.018

218 Tierney, K.B., 2016. Chemical avoidance responses of fishes. *Aquat. Toxicol.*  
219 doi:10.1016/j.aquatox.2016.02.021

220 Tomkins, P., Saaristo, M., Allinson, M., Wong, B.B.M., 2016. Exposure to an  
221 agricultural contaminant, 17 $\beta$ -trenbolone, impairs female mate choice in a  
222 freshwater fish. *Aquat. Toxicol.* 170, 365–370.  
223 doi:10.1016/j.aquatox.2015.09.019  
224