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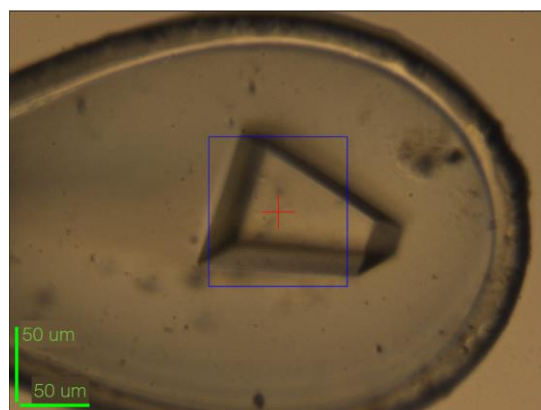
Supporting information for article:

RNA protects a nucleoprotein complex against radiation damage

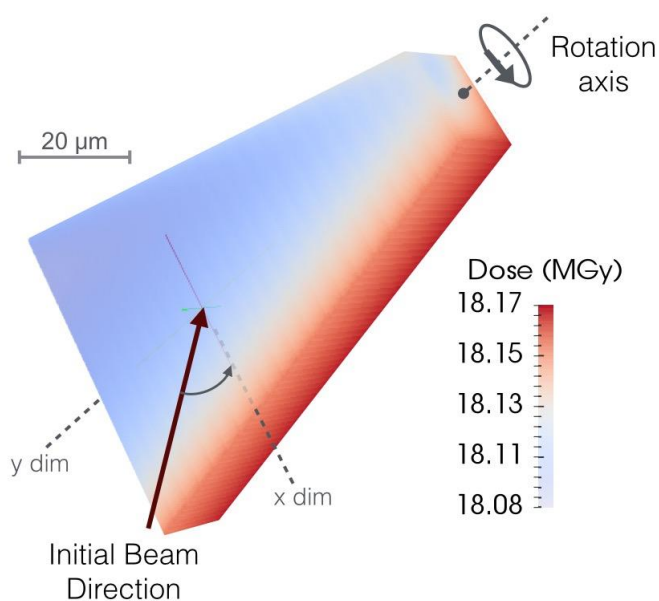
Charles S. Bury, John E. McGeehan, Alfred A. Antson, Ian Carmichael, Markus Gerstel, Mikhail B. Shevtsov and Elspeth F. Garman

Crystal	<pre> # Crystal Block Crystal Type Polyhedron PixelsPerMicron 2 AngleP 8 AngleL 90 AbsCoefCalc EXP PDB 1GTF SolventHeavyConc K 805 P 105 Mg 15 Cl 430 WIREFRAME TYPE obj Modelfile crystal_TRAP_7may2015.obj </pre>	<pre> # Polyhedron class used to implement crystal modelled in Blender # Crystal absorption coefficient to be calculated from PDB accession code (see below) # 1gtf coordinate structure from pdb (Hopcroft et al., 2002). The unit cell dimensions, number of monomers, protein residues and RNA nucleotides, and solvent volume calculated from this # Heavy elements (atomic mass larger than oxygen) within solvent calculated from crystallisation conditions (potassium phosphate, potassium glutamate KCl and MgCl2) # 3D crystal modelled with Blender # The file name of Blender crystal object </pre>
Beam	<pre> # Beam Block Beam Type TopHat Flux 5e11 Energy 13.2 Collimation Rectangular 100 100 </pre>	<pre> # Uniform beam approximation made # flux in units of photons/second # beam energy in units of keV # units of um each </pre>
Wedge	<pre> # Wedge Block Wedge 40 220 ExposureTime 180 AngularResolution 2 </pre>	<pre> # The start and stop rotation axis angles (concurrent with crystal y-axis) # The total exposure time for full wedge </pre>
Wedge	<pre> # Wedge Block 2 Wedge 40 220 ExposureTime 180 AngularResolution 2 </pre>	<pre> # Additional data collection wedges added by appending new wedge blocks here </pre>

Figure S1 RADDOS-3D (Zeldin, Gerstel, *et al.*, 2013) .txt input file for TRAP dataset 2, calculating a DWD (Diffraction-Weighted Dose (Zeldin, Brockhauser, *et al.*, 2013)) of 3.88 MGy. Crystal, beam and data collection wedge parameters are defined in three discrete input blocks. Additional data collection wedges are described by appending adding additional wedge blocks to the input file (here 2 successive 100 degree wedge of 1° images over the same angular range are simulated).

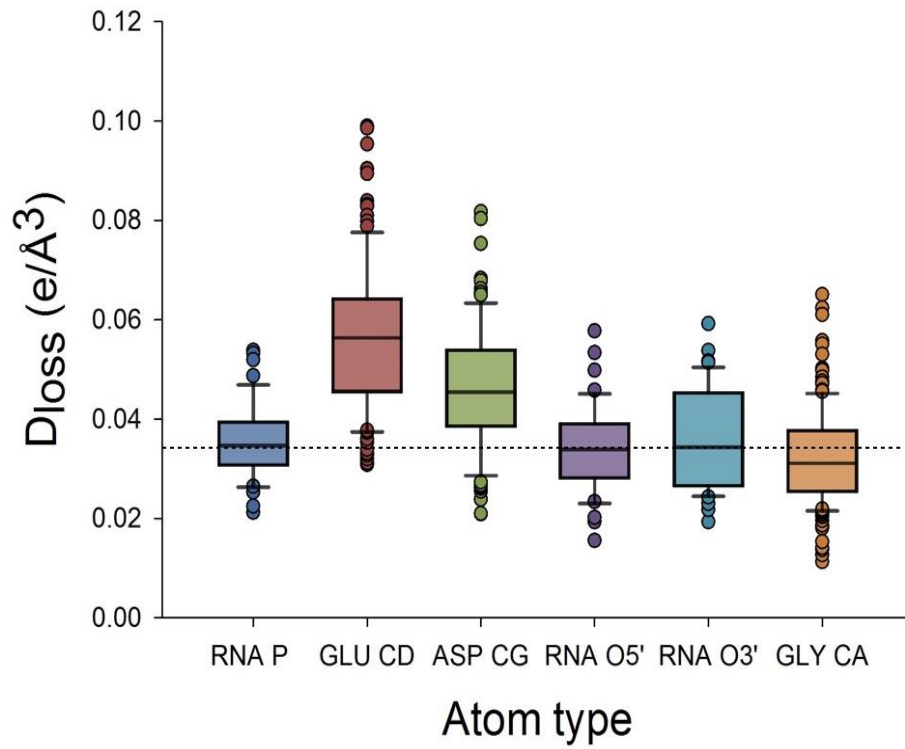


(a)

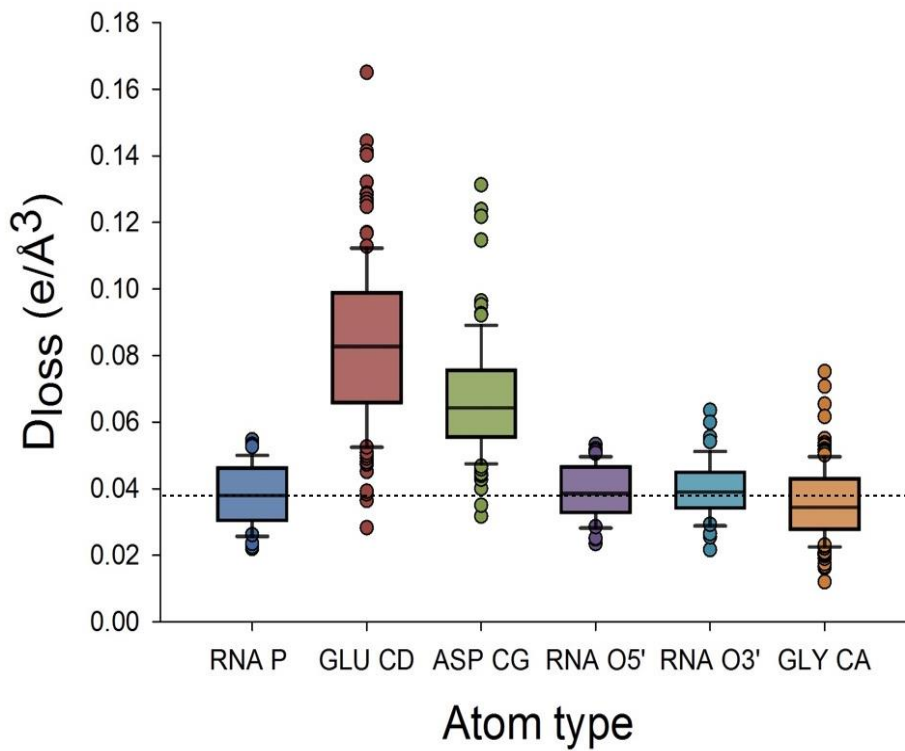


(b)

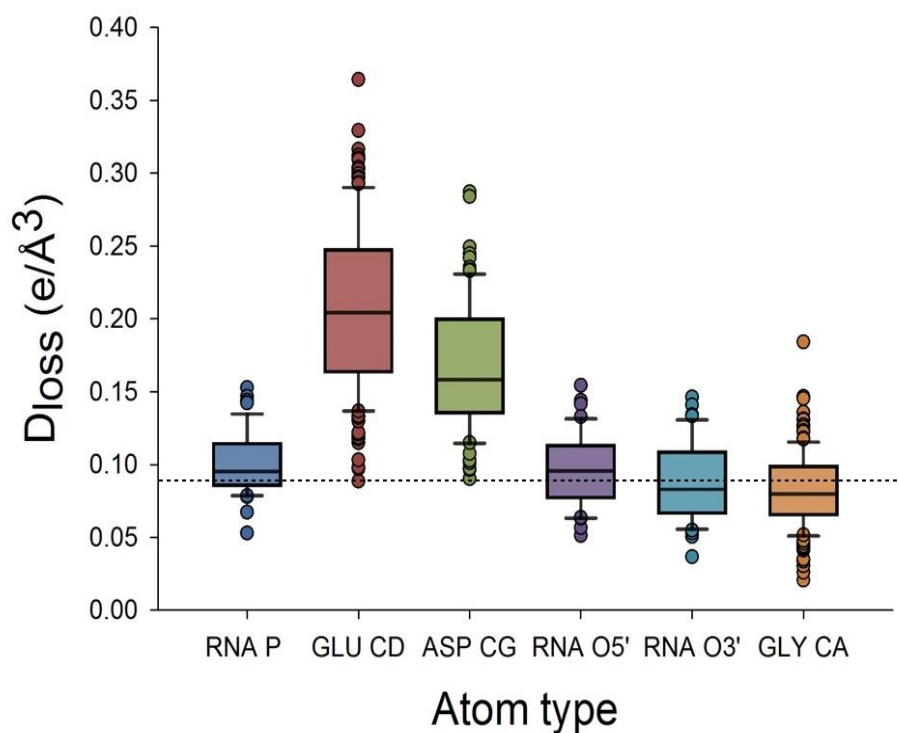
Figure S2 (a) The TRAP-RNA crystal within a rayon cryo-loop on beamline. The x and y crystal dimensions were estimated at $60\ \mu\text{m}$, $80\ \mu\text{m}$, $80\ \mu\text{m}$, $30\ \mu\text{m}$ (travelling in a clockwise direction around the crystal from the left-most side). The z dimension ($\sim\mu\text{m}$) is orientated directly into the page, perpendicular to the loop. The red crosshair indicates the beam centre, and the blue reference box shown around the crystal is $100\ \mu\text{m} \times 100\ \mu\text{m}$. (b) The dose distribution within the TRAP crystal calculated in RADDPOSE-3D for the 7th dataset (DWD: $16.7\ \text{MGy}$). The initial beam direction and rotation axis (concurrent with y -axis) are shown. At each dataset a homogenous dose distribution (to within $\sim 0.51\%$) was predicted throughout the crystal.



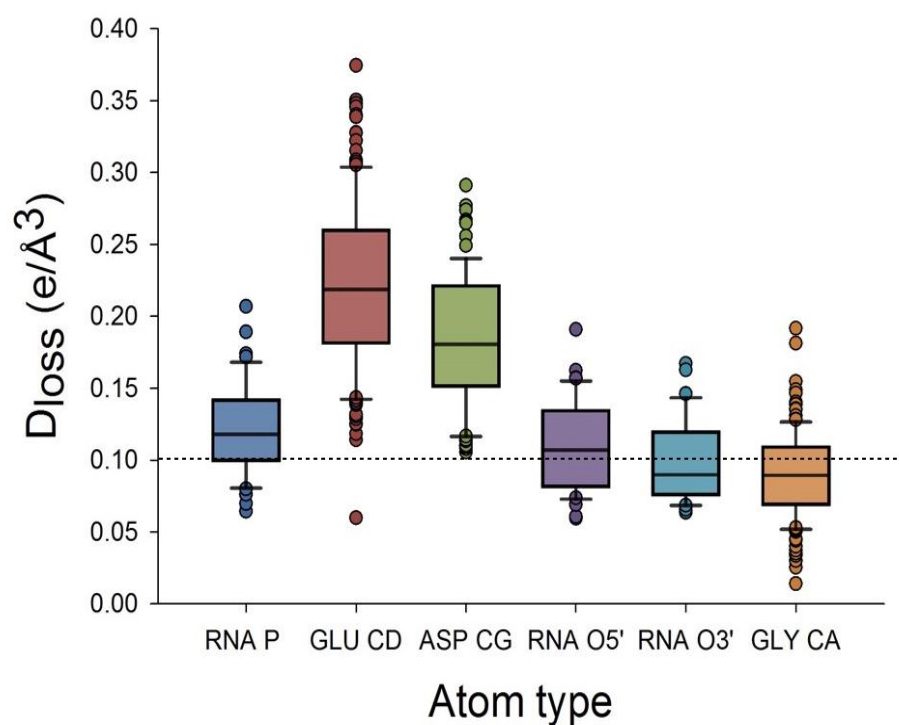
(a)



(b)



(c)



(d)

Figure S3 D_{loss} metric calculated for all RNA P, O5' and O3' atoms, and all Glu C $_{\delta}$, Asp C $_{\gamma}$ and Gly C $_{\alpha}$ atoms over 4 increasing doses (a) 3.88 MGy, (b) 6.45 MGy, (c) 21.9 MGy and (d) 25.0 MGy. The Gly C $_{\alpha}$ atom control is included as a measure of the overall D_{loss} increase with dose due to global radiation damage effects. The horizontal dashed line illustrates the average D_{loss} calculated over all

refined atoms in the TRAP complex, at each dose, as a measure of the overall rate of density disordering due to *global radiation damage*. Over the large dose range, RNA backbone atoms were determined to disorder at a rate on the same order as the overall global damage, in contrast with Glu and Asp residues, which consistently disordered at a rate above the global damage rate.

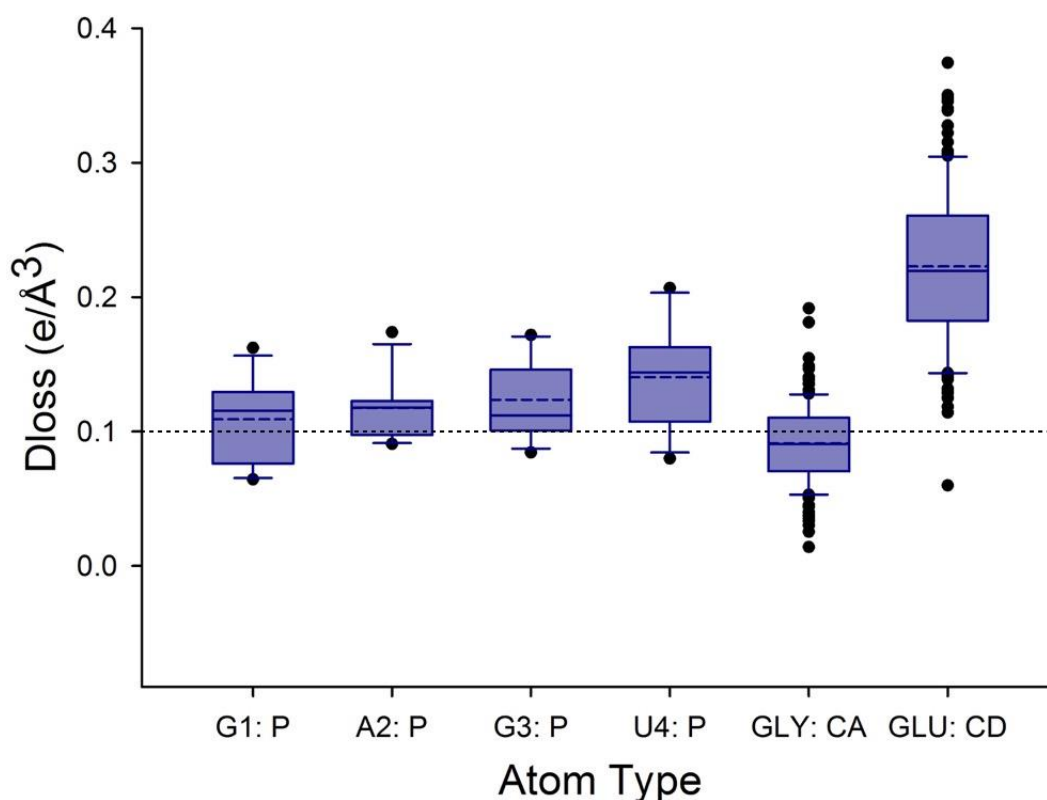


Figure S4 D_{loss} metric calculated for all phosphorus atoms for each of 4 refined nucleotide types: G1, A2, G3 and U4 at the highest investigated dose (25.0 MGy). The D_{loss} calculated for Glu C_{δ} and Gly C_{α} atoms is included for comparison. The U4 nucleotide does not interact directly with the protein, and the backbone phosphorus atom of U4 was shown to disorder on average at a rate marginally lower than that of the other refined nucleotide types (G1, A2, G3). However the U4 P atom disordering is still significantly lower than that of Glu side-chain carboxyl atoms. The horizontal dashed line illustrates the average D_{loss} calculated over all refined atoms in the TRAP complex, as a measure of the overall rate of density disordering due to *global radiation damage* effects.

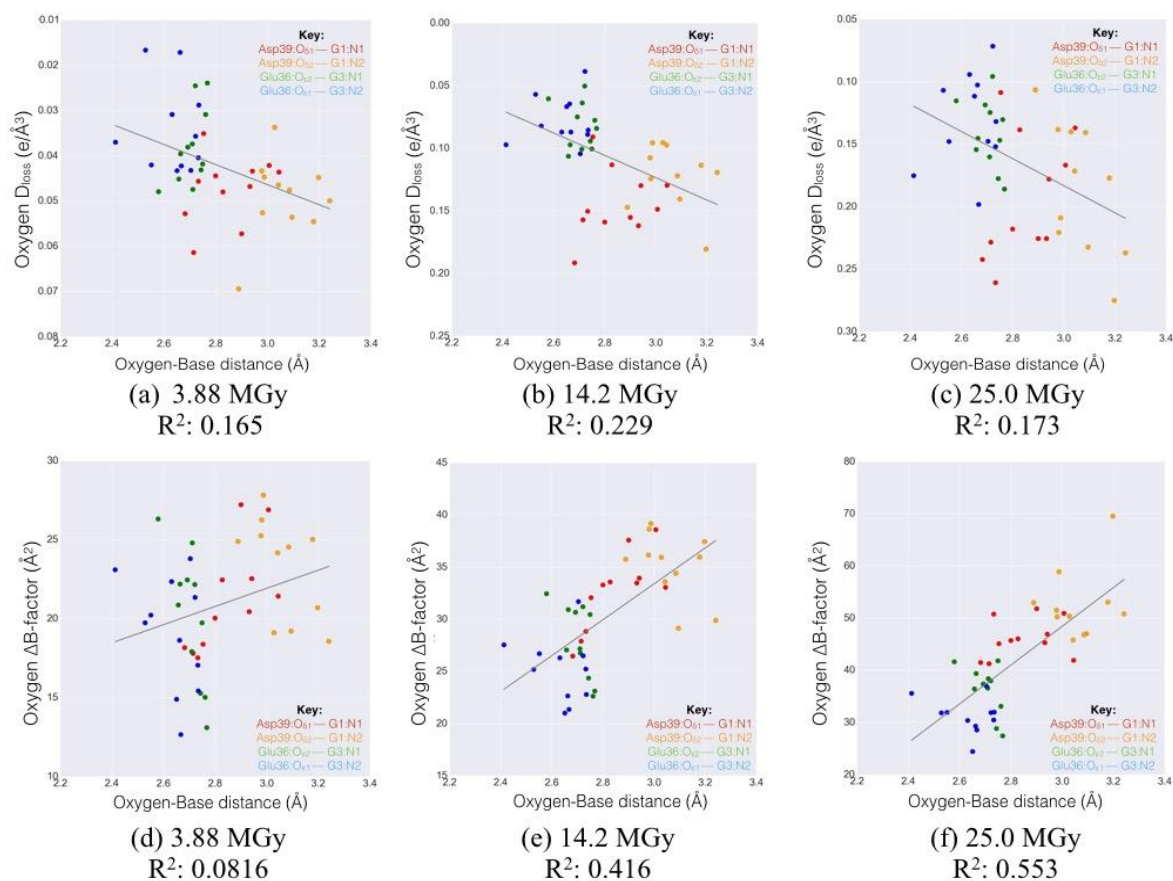


Figure S5 (a-c) D_{loss} and (d-f) ΔB -factor (change in atomic B-factor between dataset n and 1) metrics calculated for Asp 39 and Glu 36 side-chain carboxyl oxygen atoms, plotted against hydrogen bonding distance (Å) to the nearest G1 and G3 base respectively. Plots are given for 3 increasing doses between 3.88 and 25.0 MGy. Linear regression R^2 values for each scatter plot are given. With increasing dose, the ΔB -factor metric was observed to become increasingly linearly correlated with carboxyl oxygen-base hydrogen bonding length; the D_{loss} metric however did not correlate with hydrogen bond length. The Asp-39 side-chain carboxyl oxygens have been observed in previous studies to consistently make one or two distorted hydrogen bonds with the N1 and N2 of the G1 RNA base around the TRAP ring, depending on the exact RNA binding sequence (Hopcroft *et al.*, 2002, Antson *et al.*, 1999, Elliott *et al.*, 2001). For the Asp-39-G1 base interactions observed here, the mean $O_{\delta 1}$ -N1 interaction distance is 2.9 Å whereas the $O_{\delta 2}$ -N2 is consistently larger, with a mean distance of 3.1 Å. For Asp-39, only for the $O_{\delta 1}$ carboxyl oxygen were the D_{loss} dynamics statistically distinguishable between bound and non-RNA TRAP (Hotelling T-Squared Test: $O_{\delta 1}$ $p=0.019$ compared to $O_{\delta 2}$ $p=0.109$). These observations are in agreement with those made previously, in which Asp-39 was reported to interact with the G1 base only through a single distorted $O_{\delta 1}$ -N2 hydrogen bond (Hopcroft *et al.*, 2002). The Glu-36-G3 base hydrogen bond interactions were notably shorter, with mean interaction distances of 2.6 Å ($O_{\epsilon 1}$ -N2) and 2.7 Å ($O_{\epsilon 2}$ -N1). An increasing correlation with dose was observed between oxygen-base hydrogen bond length and ΔB -factor ($R^2 > 0.41$ for dose

≥ 11.6 MGy) for the individual Glu-36 and Asp-39 residues. However no direct quantitative correlation could be established between hydrogen bond length and D_{loss} (linear $R^2 < 0.23$ for all doses).

Table S1 Data processing and refinement statistics. Values in parentheses are for the highest-resolution shells. For observed F_{obs} and calculated F_{calc} structure factors, $R_{\text{work}} = \sum |F_{\text{obs}} - F_{\text{calc}}| / \sum F_{\text{obs}}$, and R_{free} is the R_{work} formula calculated from the same 5% test set of randomly selected reflections excluded throughout refinement. The current TRAP-RNA structure crystallised in space group C2 ($\alpha = \gamma = 90^\circ$). The resolution range was fixed at 63.00 - 1.98 Å throughout, with 2.01 - 1.98 Å for the outer shell for all datasets. *MolProbity* (Chen *et al.*, 2010) was used for structure validation within *phenix.refine*, giving for all structures: RMSD bond length: 0.024 Å, RMSD bond angle: 2.3°, Ramachandran favoured/outliers/allowed: 99.5/0.0/0.5%, rotamer outliers: 2.6%. The number of protein, RNA and solvent non-hydrogen atoms remained constant at 12135, 968 and 743 respectively for all dataset numbers, due to the rigid body refinement of higher dose datasets.

Dataset	1	2	3	4	5	6	7	8	9	10
DWD (MGy)	1.31	3.88	6.45	9.02	11.58	14.15	16.72	19.29	21.86	24.98
Data processing										
Cell dimensions										
a (Å)	140.90	141.00	141.04	141.04	141.12	141.14	141.16	141.19	141.21	141.23
b (Å)	110.89	110.98	111.02	111.05	111.08	111.10	111.13	111.17	111.16	111.16
c (Å)	137.81	137.93	137.99	138.05	138.09	138.15	138.18	138.21	138.28	138.29
β (°)	117.41	117.41	117.41	117.40	117.40	117.39	117.39	117.39	117.39	117.37
# observs.	473080	471462	472091	472756	473225	476499	477213	477019	475519	472868

Dataset	1	2	3	4	5	6	7	8	9	10
# Unique reflc.	129942	130202	130317	130454	130599	130738	130870	130920	130982	130840
Multiplicity	3.6 (3.5)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)	3.6 (3.4)
Completeness (%)	99.3	99.3	99.3	99.3	99.3	99.3	99.3	99.3	99.3	99.1
R _{merge}	0.08 (0.70)	0.08 (0.78)	0.08 (0.86)	0.09 (1.02)	0.09 (1.13)	0.10 (1.31)	0.10 (1.50)	0.11 (1.68)	0.11 (2.07)	0.11 (1.98)
Mean(I)/sd(I)	9.6 (1.5)	9.4 (1.4)	9.3 (1.2)	8.8 (1.1)	8.4 (1.0)	8.1 (0.8)	7.4 (0.7)	7 (0.6)	7 (0.5)	7.2 (0.5)
I _n /I ₁	1	0.905	0.865	0.831	0.776	0.738	0.676	0.638	0.625	0.854
Refinement										
R _{work}	0.192	0.209	0.209	0.212	0.216	0.220	0.225	0.230	0.234	0.240
R _{free}	0.231	0.244	0.243	0.245	0.248	0.252	0.256	0.263	0.264	0.271
Mean B-factor (Å ²)	33.78	34.50	36.04	37.22	38.86	40.35	42.89	44.70	47.71	47.58

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