

# Exercise performance in acute and chronic cold exposure

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## Abstract

This review focuses on the suppression of physical performance in a cold environment and the underlying physiological mechanisms. There are many situations where humans have to perform physical activities in a cold environment. Cold environments often limit exercise and working performance by impairing functions such as force production, velocity, power and manual dexterity. A muscle temperature of around 27°C is assumed to be a critical temperature below which maximal voluntary isometric force starts to decrease. The endurance time of submaximal isometric contractions peak at muscle temperatures of 27 to 28°C and decrease rapidly above and below these temperatures. Dynamic exercise performance, especially fast velocity movement, is generally more disturbed by cooling than isometric contractions. Additionally, the effect of cold adaptation on exercise performance, and the potential related mechanisms are summarized here based on a limited number of studies. Since the involuntary muscle contraction of shivering disturbs fine motor control, habituation of shivering, which is an example of cold adaptation, potentially improves exercise performance. Higher hand skin temperatures, induced by greater cold induced vasodilatation after local cold adaptation, could improve manual dexterity. Since there have been few studies testing the effect of cold adaptation on exercise performance in a cold environment, further studies seem warranted.

## Introduction

Humans are exposed daily to various climatic conditions, from mild to extreme hot and cold, and sometimes experience meteorological disasters like cold snaps, heavy snow fall, flooding and tsunamis, which results in cold-associated, sometimes fatal, health conditions. In the case of accidental cold water immersion, the initial drop of skin temperature induces “cold shock” responses including hyperventilation, gasping and tachycardia<sup>1</sup>). These responses peak in the first 30 seconds, and are probably responsible for the majority of drowning deaths<sup>1</sup>). Following skin cooling, incapacitation caused by a reduction in temperature of superficial nerves and muscle, can impair performance and even result in death before hypothermia. The impairment of muscle performance of the hypothermic skeletal muscle induces swimming failure<sup>2</sup>) and drowning. Even in recreational or competitive sports events in a cold environment like skiing, snowboarding, diving and long distance swimming, exercise performance can be restricted by the cold. In addition, decreased performance due to a reduction of the superficial tissue temperature is commonly experienced by workers in cold workplaces like the food industry, cold storage, fisheries, forest industry, Coast Guard and military, especially in cold countries in the winter season<sup>3,4</sup>).

There are many situations where humans have to perform physical activities in a cold environment. Cold environments often limit exercise and work performance by affecting force production, velocity, power and manual dexterity<sup>3,5</sup>). Chronic cold exposure and repetitive work might lead to cold-related health problems like musculoskeletal pain, Raynaud's phenomenon (recurrent vasospasm in the fingers and toes), and, on some occasions, non-freezing cold injuries and frostbite<sup>4,6</sup>). On the other hand, the benefits of cooling on exercise performance have been studied especially for prolonging endurance exercise in the heat, for example, pre-cooling or cooling between exercise sessions<sup>7-9</sup>). The major focus of this review is the cold-induced impairment of exercise performance, excluding these possible positive effects. Additionally, the greater cold-induced vasodilatation (CIVD) response observed in people working daily in cold environments<sup>10-12</sup>), has been suggested to be beneficial for preventing cold injuries and improving manual dexterity in the cold<sup>13</sup>).

This review article focuses on the impairment of physical performance in cold environments and underlying physiological mechanisms. Additionally, the effect of cold adaptation on exercise performance and related potential mechanisms are summarized based on a limited number of studies.

## **Exercise performance in a cold environment**

The impairment of exercise performance in hypothermic skeletal muscle has been studied and well summarized in several review articles<sup>5,14-16</sup>). In this review, the exercise performance of static isometric contraction, dynamic exercise, and the more practical swimming exercise are summarized below.

### **Isometric contraction**

The classic studies of Clarke et al.<sup>17</sup>) reported a small or no effect of muscular temperature on maximal voluntary contraction (MVC) during isometric handgrip within the muscle temperature ( $T_m$ ) range of 27 to 40°C, but a decrease of 60% when  $T_m$  reached around 20°C. Similarly, in the first dorsal interosseous muscle, MVC was relatively constant, or even greater, within the range of 25 to 35°C, but decreased by about 30% when  $T_m$  was 12 to 15°C. Others have reported a small decrease of MVC during isometric knee extension (2.1% MVC per °C muscle temperature reduction) within the range of 30 to 39°C<sup>18</sup>). Several studies reported a reduction in MVC when  $T_m$  dropped below 27°C<sup>17,19</sup>). While there is some variation between studies, a muscle temperature of around 27°C is assumed to be a critical temperature for performing maximal isometric voluntary contraction<sup>5,16</sup>).

Meanwhile, endurance performance follows a bellshaped pattern with muscle temperature. The endurance time of submaximal isometric contraction peaks at a  $T_m$  of 27 to 28°C and decreases rapidly above and below these temperatures<sup>17,20</sup>). When  $T_m$ , measured at a 2 cm depth of the brachioradialis, was decreased

to 23°C or elevated to 38°C, the duration of sustained hand grip contraction (33% MVC) was shortened by around 60% of the peak observed at 27°C<sup>17</sup>). It was suggested that at a  $T_m$  below 27°C superficial muscle fibers do not contract well, and fewer fibers located more centrally have to generate the same force. When  $T_m$  increased above 27°C, the increase in metabolic rate caused earlier fatigue<sup>17</sup>).

### **Dynamic exercise**

Bergh and Ekblom<sup>18</sup>) measured the peak torque of knee extension at the different angular velocities of 0° (isometric), 90° and 180°·sec<sup>-1</sup>, within a  $T_m$  range of 30 to 39°C. The decrease in peak torque per °C reduction of  $T_m$  was greater in dynamic conditions (4.7%·°C<sup>-1</sup> at 90°·sec<sup>-1</sup> and 4.9%·°C<sup>-1</sup> at 180°·sec<sup>-1</sup>) than in an isometric condition (2.1%·°C<sup>-1</sup> at 0°·sec<sup>-1</sup>). It was suggested that dynamic exercise performance, especially fast velocity movement, is generally more disturbed by cooling than isometric contractions<sup>5,14,18</sup>).

Maximal 20-sec sprint efforts at 95 revolutions per min on an isokinetic cycle ergometer were tested under different muscle temperature conditions following leg immersions<sup>21</sup>). When  $T_m$  was reduced to 31.9 and 29.0°C, the maximal peak force was reduced by 12 and 21%, respectively, compared to a non-cooling condition at a  $T_m$  of 36.6°C; this corresponded to a 2-3% reduction per °C fall in  $T_m$ <sup>21</sup>). The height of a maximal vertical jump was decreased with a  $T_m$  reduction within the range of 30 to 39°C at the rate of 4.2%·°C<sup>-1</sup><sup>18</sup>). Similarly, the flight time, force production during shortening phase and take-off velocity of maximal drop jump were decreased in a dose-dependent manner with degree of cooling<sup>22</sup>). It was suggested that very fast movements like the drop jump are especially susceptible to cooling<sup>14</sup>). Sargeant<sup>21</sup>) tested exercise performance at different pedaling velocities, and found a  $T_m$  dependence on the optimum pedaling rate for maximal power production. This velocity dependence of performance in cold was also reported in a ball throwing exercise<sup>23</sup>); cold-induced impairment was greater with lighter balls (faster movement) than with heavier balls (slower movement).

Time to exhaustion during short-term intense leg cycling exercise was reduced by 38% when leg muscles were cooled to 29°C, compared to a warm condition of a  $T_m$  of 34°C<sup>24</sup>). Similarly, the endurance performance of intense dynamic exercise was impaired by cooling<sup>25,26</sup>).

### **Swimming**

Swimming in cold water can significantly impair exercise performance; the greater thermal conductance of water quickly decreases body temperature. Swimming needs coordination of the dynamic movement of different body parts, proprioceptive feedback and tactile sensation for perceiving motion in the water. These functions are also impaired by cooling. In the well-known studies on the metabolic response to swimming in varying water temperatures (18, 26 and 33°C), greater oxygen uptake for a given submaximal swimming speed (less efficient swimming) was observed in colder water<sup>27,28</sup>). It was suggested that the

additional energy expenditure was attributable to shivering and lower mechanical efficiency due to impaired neuromuscular function in cold limbs. Additionally, lower oxygen uptake during maximal swimming was observed in lean participants<sup>27,28</sup>). The details of swimming failure (stroke parameters and efficiency) were analyzed during breaststroke swimming in cold water at 10, 18 and 25°C, for a maximum of 90 min<sup>2</sup>). A more frequent stroke rate (number of strokes in a minute) and shorter stroke length (distance swum per each stroke) were observed in 10°C water than in warmer water. Impaired swimming performance can partly explain the greater oxygen consumption and diminished swimming efficiency (meters swum per liter of oxygen consumption) in 10°C water.

### **Mechanism of performance impairment in a cold environment**

The reduction in hypothermic skeletal muscle performance described is, in part, attributable to slowing of the nerve conduction velocity<sup>29</sup>). Reduction of the muscle contraction velocity is partly explained by slowed adenosine triphosphate (ATP) utilization<sup>30,31</sup>), slowed Ca<sup>2+</sup> release and uptake from the sarcoplasmic reticulum<sup>32,33</sup>). In this section, the potential mechanisms of impairment of exercise performance in a cold environment are summarized based on studies focusing on neuromuscular functions and muscle metabolism.

### **Neuromuscular function**

The neuromuscular function of hypothermic skeletal muscle has been studied using electromyography (EMG). Several studies have reported decreased amplitude of EMG due to cooling<sup>34-36</sup>), while others have reported increased amplitude<sup>37,38</sup>). The discrepancy could be explained by different exercise types or cooling procedures. During a brief biceps contraction at 30% maximum voluntary contraction (MVC), greater amplitude was observed when the upper arm was moderately cooled by a 20°C water circulating cuff, whereas, the amplitude was reduced when water temperature was reduced to 0°C<sup>39</sup>). When muscle was cooled from the skin surface, relatively cooler superficial muscle fibers did not contract well, and warmer fibers located more centrally with the muscle compensated for the required force. Thus, the reduction of the amplitude of surface EMG might reflect the lower activity of superficial muscle fibers<sup>40</sup>). On the other hand, the increase of amplitude may indicate that more muscle fibers are recruited to maintain the given work load level<sup>41</sup>). It has been reported that faster muscle fibers in swimming carp are recruited at relatively lower velocity in low temperature conditions to maintain swimming speed<sup>42,43</sup>). The maximal shortening velocity of rat fast (extensor digitorum longus) and slow (soleus) twitch fibers in vitro has been examined at temperatures between 35 and 10°C<sup>44</sup>). It was found that slow twitch fibers showed a greater decrease in the shortening velocity per °C reduction of  $T_m$  than fast twitch fiber. Because of the greater cold sensitivity and lower power output of slow twitch fibers in a cold environment, less cold sensitive and more powerful fast twitch fibers are recruited earlier. Therefore, to generate the muscle

power to maintain work load, a greater number of fast twitch fibers need to be recruited in cold compared to a normothermic condition. This results in a greater amplitude of EMG.

A shift in the EMG frequency to lower frequencies with lower muscle temperatures has been reported more uniformly<sup>22,34,35,37,39,45</sup>). The shift has been connected with the decrease in muscle conduction velocity in the cold<sup>39,46</sup>). Since the reduction of conduction velocity with muscle cooling paralleled the reduction in the median of EMG frequency, the shift to lower frequencies would reflect a change in muscle action potential conduction velocity<sup>35</sup>). Since a similar reduction of EMG frequency was observed in fatigued muscle, the mechanism of the lower EMG frequency in cold was explained with the fatigue-induced 'muscular wisdom' hypothesis<sup>15</sup>). Marsden et al.<sup>47</sup>) developed the 'muscular wisdom hypothesis' that was a phenomenon of the decrease in the motor unit discharge rate during continuous muscle contraction for minimizing fatigue. Since fast twitch fibers, which are less involved in endurance activity, are recruited earlier in the cold<sup>42,43</sup>), it is suggested that optimizing the firing frequency is beneficial for fatigue resistance<sup>15</sup>).

There are only a few studies examining the co-contraction of the agonist and antagonist muscle pair after cooling<sup>22,23</sup>). During the concentric phase of muscle contraction, the EMG activity of the antagonist muscle is increased by cooling, whereas activity of the agonist is suppressed. This phenomenon called the 'braking effect' has been suggested as one reason for the impairment of exercise performance<sup>14,16</sup>). Similarly, involuntary shivering of the antagonist muscles can disturb control of motor activity in a cold environment<sup>48-50</sup>). Hong and Nadel<sup>51</sup>) tested the effect of exercise intensity on the shivering of neck muscles which were not involved in exercise. The slope relationship of neck muscle EMG activity to esophagus temperature was suppressed by increasing exercise intensity.

### **Muscle metabolism**

Abramson et al.<sup>52</sup>) examined resting local oxygen uptake and blood flow in human forearm at a  $T_m$  within the range of 25 to 40°C. The oxygen uptake was calculated using the Fick principle on blood samples from the forearm vein, and forearm blood flow was measured by plethysmography. Lower local oxygen uptake and forearm blood flow was seen in cold muscle. The suppression of muscle metabolism in a cold environment could partly be explained by a reduction of oxygen supply by blood flow restriction in the cold<sup>53-55</sup>). Thorsson et al.<sup>53</sup>) reported a reduction of intramuscular blood flow (<sup>133</sup>Xe clearance technique) after local cooling of the quadriceps by applying cold packs. Rennie et al.<sup>55</sup>) showed a similar reduction of muscle blood flow during exercise in cold water.

There have been large numbers of studies measuring oxygen uptake in the lungs to evaluate the effect of cold muscle temperature on the metabolic response. Temperature dependence on oxygen uptake kinetics

at the start of exercise has been studied. Shiojiri et al.<sup>56)</sup> found a significantly greater time constant (the duration to the end of phase 1, defined as inflection points in respiratory exchange ratio, end-tidal PO<sub>2</sub> and end-tidal PCO<sub>2</sub><sup>57)</sup>) after onset of moderate (50 W) cycle exercise under a cold muscle condition (vastus lateralis  $T_m = 30.2^\circ\text{C}$ ) than in a neutral condition ( $T_m = 36.8^\circ\text{C}$ ). On the other hand, Ishii et al.<sup>58)</sup> reported no difference between cold (vastus lateralis  $T_m = 28.0^\circ\text{C}$ ) and neutral ( $T_m = 35.5^\circ\text{C}$ ) conditions in oxygen uptake kinetics (half-times of oxygen uptake kinetics) at the onset of cycle exercise (75 W and 125 W). In the cold conditions of both studies, rectal temperature was also decreased before starting exercise, which would induce shivering. Significantly greater oxygen consumption before starting exercise was observed in the cold conditions of the study by Ishii et al.<sup>58)</sup>, whereas Shiojiri et al.<sup>56)</sup> reported similar resting oxygen consumption in both temperature conditions. Thus, in the work of Ishii et al.<sup>58)</sup> a deviation in the initial level of oxygen consumption due to shivering may have masked the difference in oxygen kinetics between muscle temperature conditions. Recently, the kinetics of intracellular oxygen pressure (PO<sub>2</sub>) following the onset of contraction was evaluated in isolated single *Xenopus* skeletal muscle fibers maintained at different temperatures<sup>59)</sup>; a greater time constant and delayed onset for the decline of intracellular PO<sub>2</sub> was observed in cold muscle ( $T_m = 15.4^\circ\text{C}$ ) than in muscle kept in a neutral ( $T_m = 20.5^\circ\text{C}$ ) or hot condition ( $T_m = 25.9^\circ\text{C}$ ).

It was suggested that developing techniques of measuring local oxygen uptake was essential in order to study human muscle metabolism over a large range of  $T_m$ <sup>60)</sup>. Recently, several studies directly measured the temperature dependence of muscle metabolism using near infrared spectroscopy (NIRS)<sup>61-63)</sup>. It was reported that oxygen consumption of human forearm at rest and during 4% MVC isometric handgrip (by measuring the slope of deoxyhemoglobin change during 20-sec arterial occlusion) decreased as a function of reduction in  $T_m$  from 36 to 26°C<sup>61)</sup>. Using the NIRS technique, a significant reduction of total hemoglobin level and a tendency for lower muscle oxygenation were observed in resting human ankle dorsiflexor muscles cooled from the skin surface<sup>63)</sup>. Hom et al.<sup>62)</sup> also reported a decrease in resting muscle oxygen saturation and total hemoglobin levels after 1-hour cooling with an ice bag.

More recently, diffusion-weighted magnetic resonance images were used to evaluate the effect of cooling on intramuscular water movement, including water diffusion and microvascular circulation in the capillary network<sup>9,64,65)</sup>. It was reported that resting intramuscular water diffusion and perfusion were decreased by local cooling<sup>64)</sup>, and the increased intramuscular water movement after exercise showed greater recovery after local cooling treatments than in a non-cooling condition<sup>65)</sup>. This magnetic resonance technique could be applied to directly evaluate microscopic water movement in hypothermic skeletal muscle, which is associated with muscle metabolism.

## **Cold adaptation and exercise performance**

Cold adaptation of human thermoregulatory response has been studied extensively, and there have been several categorizations based on the differences in thermoregulatory response following a period of adaptation<sup>66-70</sup>). In this section, the effect of repeated cold exposure on exercise performance in a cold environment and potential related mechanisms are summarized based on a limited number of studies.

### **Habituation of shivering**

It has been reported that shivering can disturb fine motor control because of the co-contraction of the agonist and antagonist muscles with the superimposition of shivering on exercise in a cold environment<sup>48-50</sup>). Thus, the habituation of shivering (less shivering response after cold adaptation) could improve fine motor control in the cold. There are a lot of studies reporting the habituation of shivering response after repeated exposure to a cold environment<sup>71-73</sup>). However, only a limited number of studies tested the influence of adaptation on exercise performance.

Makinen et al.<sup>74</sup>) found a significant reduction of postural sway with a significant increase in mean skin temperature (0.4°C) over 10 days repeated exposure to 10°C air for 2 hours per day. However, since the postural sway at 10 and 25°C ambient temperature conditions improved in parallel, it was concluded that this improvement was attributed to motor skill learning and the repeated cooling had no effect on postural control<sup>74</sup>). During cold exposure to 10°C air, muscle tone measured by EMG was increased by 140-260% (visible shivering in some participants); but no habituation was observed over the 10-day exposure period. The temperature condition of 10°C air might not have been sufficient to habituate shivering. If the environmental condition was much colder, inducing strong shivering, an improvement of neuromuscular performance might be observed with the habituation of shivering. A recent study also reported that athletes who practiced in cold weather had no improvement in their manual dexterity, despite the smaller reduction in their finger skin temperature during cold exposure to 5°C air<sup>75</sup>).

Research has examined neuromuscular adaptation after repeated 2-min whole-body exposure to extremely cold air (-110°C) for 3 months<sup>76</sup>). At the first trial before the adaptation, the flight time of a drop jump was significantly shortened by a single 2-min cold exposure, but this impairment of performance disappeared after the repeated exposure to cold air. Based on the simultaneously measured EMG data, the improvement in performance was explained by the pronounced increase in activity of the agonist muscle (gastrocnemius medialis) during the shortening phase of the drop jump after repeated cold exposure; no change was observed in the antagonist (tibialis anterior). The diminished co-contraction of the agonist and antagonist muscles<sup>48</sup>), or braking effect, by the increased antagonist muscle activity<sup>22,23</sup>) in a cold environment, might improve the drop jump performance. Additionally, maximal voluntary contraction (MVC) of isometric wrist flexion was evaluated with the agonist and antagonist EMG analysis. The averaged

EMG activity of both the agonist and antagonist muscles was significantly increased following the 2-min single cold exposure after the 3-month repeated exposure to cold. Westerlund et al.<sup>76)</sup> commented that an increase in EMG activity tended to be greater in the agonist and less in the antagonist, which is similar to the reduced co-contraction during drop jump after repeated cold exposure. However, wrist flexion exercise performance did not change significantly either following a single 2-min cold exposure or after repeated exposures. The discrepancy between the results was suggested to be due to the difference of muscle contraction type (isometric or dynamic), since the maximal isometric force was relatively stable when the muscle temperature was over 27°C<sup>17)</sup> and the impairment of muscle performance in the cold was greater during dynamic than static exercise<sup>5,14,26)</sup>.

### **Enhancement of cold induced vasodilatation**

During cold-water immersion of extremities, higher finger skin temperature, more rapid cold induced vasodilatation (CIVD) and enhanced blood flow to the extremities are generally, but not always<sup>77-80)</sup>, observed in populations living and working in a cold environment<sup>10-12)</sup> and also after repeated cold exposure in laboratory controlled studies<sup>81,82)</sup>. The greater CIVD response was suggested to be beneficial for improving manual dexterity and tactile sensitivity in the cold<sup>13)</sup>. Manual dexterity is affected by finger skin temperature<sup>83)</sup> or finger blood flow<sup>84)</sup> depending on the temperature and duration of the task. Krog et al.<sup>10)</sup> reported that the impairment of grip strength and finger tapping speed following 30-min hand immersion in 0°C water tended to be smaller in Norwegian Lapps and fishermen than in control groups. In their study, cold-adapted groups showed a significantly rapid onset of CIVD, however, no difference was observed in maximum and minimal finger skin temperature compared to a control group. Thus, it was not clear whether the manual dexterity of cold-adapted populations were affected by habituation of local vasoconstrictor response.

In a series of laboratory controlled studies<sup>79,81,85)</sup>, local cold acclimation on neuromuscular functions was tested after repeated hand immersion in 8°C water for 30 min, 5 days a week for 2 to 3 weeks. No improvement in manual dexterity, grip strength, voluntary and evoked twitch force of the first dorsal interosseus muscle was observed after cold immersion<sup>79,81,85)</sup>, even with a greater CIVD response after the repeated cold water exposure<sup>81)</sup>. Concerning these observations, there seems to be no significant effect of repeated local cooling on exercise performance in cold, at least in the conditions that were tested in these studies<sup>79,81,85)</sup>.

### **Muscle metabolism**

Savourey et al.<sup>86)</sup> examined the effect of repeated local cold immersion of hand and forearm on skeletal muscle metabolism. After local cold adaptation by repeated 5-min water immersion (5°C) of the arm twice a day, 5 days a week for 2 months, finger skin temperature was kept higher during the immersion, but no

adaptive change of muscle metabolism, measured by  $^{31}\text{P}$  nuclear magnetic resonance, was observed during water immersion and during 10% MVC handgrip exercise after the immersion. Since the duration of each immersion (5 min) was too short to induce a significant reduction of  $T_m$  and muscle metabolism, the experimental protocol and small number of subjects ( $n=5$ ) may have been insufficient to induce adaptive change in muscle metabolism or identify any such change.

Muller et al.<sup>87</sup>) reported that cold weather athletes, who practiced American football in cold air ( $0^\circ\text{C}$  on average), showed a greater exercise economy (smaller oxygen uptake for a given submaximal workload) during bicycle exercise in  $5^\circ\text{C}$  air compared to a physically active control group. It was speculated, without supporting data, that improved buffering of lactate or utilization of different muscle fiber types could be a mechanism underpinning the greater exercise economy. The following are potential mechanisms which might induce cold adaptive changes in muscle metabolism.

The distribution of muscle fiber type was assessed in Korean diving women who routinely exposed themselves to cold water<sup>88</sup>). Divers had a greater percentage of type IIx (fast glycolytic) and lower proportion of type IIa (fast oxidative glycolytic) fibers in the vastus lateralis than physically active controls, whereas, no group difference was observed in the percentage of type I (slow oxidative) fibers. This result suggested that repeated cold-water immersion might induce the shift of type II muscle fibers to the faster subgroup. It was probably because the faster types of muscle fibers were recruited at a relatively lower velocity in low temperature conditions to maintain the required activity<sup>42,43</sup>). A similar shift in fiber type from type I to type IIa fibers was observed in rat soleus muscle (predominantly Type I) after intermittent cold exposure<sup>89</sup>). Since animal studies reported that the shift of muscle fiber type composition after repeated cold exposure was specific to the predominant fiber type<sup>89,90</sup>), variation in the cold adaptive change in the different muscle groups should be considered.

Another potential mechanism for improving muscle metabolism after cold adaptation is an increment of oxygen delivery caused by an increase in capillary density. Bae et al.<sup>88</sup>) reported that Korean diving women had a significantly greater capillary density and number of capillaries per fiber of the vastus lateralis compared to that of an active control group. In animal studies, chronic cold exposure has been reported to induce a growth of capillary density and/or the capillary to fiber ratio in rats<sup>91,92</sup>) and in guinea pigs<sup>93</sup>). Deveci and Egginton<sup>94</sup>) considered muscle group specificity for angiogenesis (growth of capillary) after repeated cold exposure. Since a significant increase in the capillary to fiber ratio was observed only in the soleus of rats, and not in the tibialis anterior, the greater oxidative capacity and muscle activity was suggested to be a key factor controlling cold-induced angiogenesis<sup>94</sup>). An increased oxygen supply to muscle tissues by microvascular remodeling might be a potential mechanism for improving muscle metabolism.

Recently, several research groups have focused on the adaptation of non-shivering thermogenesis (NST) in human brown adipose tissue (BAT) after repeated cold exposure<sup>95-98</sup>). Skeletal muscle potentially contributes towards NST, since it constitutes up to 40-50% of total body mass and contains large numbers of mitochondria. There was a significant positive relationship between an increase in total daily energy expenditure during mild cold exposure (16°C air for 48 hours) and an increase in mitochondrial uncoupling (state 4 respiration) of isolated human skeletal muscle biopsies taken after cold exposure<sup>99,100</sup>). However, a more recent study reported no significant contribution of human skeletal muscle mitochondrial uncoupling to an increase of NST after repeated mild cold exposure<sup>97</sup>). Concerning these observations, acute mild cold exposure would activate mitochondrial uncoupling in skeletal muscle, though, it seems to be insufficient to induce an adaptive change that enhances NST in the skeletal muscle after chronic cold exposure.

Further investigation is required to clarify the effect of chronic cold exposure on adaptation in exercise performance and physiological mechanisms.

## **Conclusion**

This article reviewed the impairment of physical performance in a cold environment and underlying physiological mechanisms. A muscle temperature of around 27°C is thought to be a critical temperature below which maximal isometric voluntary force starts to decrease; the endurance time of submaximal isometric contraction peaks at a muscle temperature of 27 to 28°C and is decreased below and above the temperature. Dynamic exercise performance is generally more disturbed by cooling than isometric contractions.

Decreased nerve conduction velocity and slowed ATP utilization could be potential mechanisms for impaired exercise performance. When skeletal muscle is cooled, a shift in the EMG frequency to lower frequencies has been uniformly reported, whereas the amplitude of EMG decreased or increased depending on the study. More muscle fibers, especially the faster type, are recruited in low temperature conditions to maintain a given workload. Several studies reported a reduction in resting muscle metabolism in the cold, but studies of exercising muscle in the cold are scarce.

Habituation of shivering and enhanced CIVD after repeated cold exposure could be hypothesized to improve exercise performance in a cold environment. Further investigation has to be carried out to clarify the effect of chronic cold exposure on adaptation in exercise performance and physiological mechanisms.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this article.

## References

- 1) Tipton MJ. 1989. The initial responses to cold-water immersion in man. *Clin Sci* 77: 581-588.
- 2) Tipton MJ, Eglin C, Gennser M and Golden F. 1999. Immersion deaths and deterioration in swimming performance in cold water. *The Lancet* 354: 626-629.
- 3) Makinen TM. 2007. Human cold exposure, adaptation, and performance in high latitude environments. *Am J Hum Biol* 19: 155-164.
- 4) Holmer I, Hassi J, Ikaheimo TM and Jaakkola JJK. 2012. Cold stress: effect on performance and health. *In: Patty's Toxicology* (Bingham E, Cohrssen B, eds.), 6th ed: 11-36, John Wiley & Sons, Inc.
- 5) Heus R, Daanen HA and Havenith G. 1995. Physiological criteria for functioning of hands in the cold: a review. *Appl Ergon* 26: 5-13.
- 6) Makinen TM and Hassi J. 2009. Health problems in cold work. *Ind Health* 47: 207-220.
- 7) Hayashi K, Honda Y, Ogawa T, Wada H, Kondo N and Nishiyasu T. 2004. Effects of brief leg cooling after moderate exercise on cardiorespiratory responses to subsequent exercise in the heat. *Eur J Appl Physiol* 92: 414-420.
- 8) Siegel R, Mate J, Watson G, Nosaka K and Laursen PB. 2012. Pre-cooling with ice slurry ingestion leads to similar run times to exhaustion in the heat as cold water immersion. *J Sports Sci* 30: 155-165.
- 9) Yanagisawa O, Otsuka S and Fukubayashi T. 2014. Effect of cooling during inter-exercise periods on subsequent intramuscular water movement and muscle performance. *Scand J Med Sci Sports* 24: 11-17.
- 10) Krog J, Folkow B, Fox RH and Andersen KL. 1960. Hand circulation in the cold of Lapps and North Norwegian fisherman. *J Appl Physiol* 15: 654-658.
- 11) Leblanc J, Hildes JA and Heroux O. 1960. Tolerance of Gaspé fishermen to cold water. *J Appl Physiol* 15: 1031-1034.
- 12) Tanaka M. 1971. Experimental studies on human reaction to cold. Vascular hunting reaction of workers to cold. *The Bulletin of Tokyo Medical and Dental University* 18: 169-177.
- 13) Daanen HA. 2003. Finger cold-induced vasodilation: a review. *Eur J Appl Physiol* 89: 411-426.
- 14) Oksa J. 2002. Neuromuscular performance limitations in cold. *Int J Circumpolar Health* 61: 154-162.
- 15) Drinkwater E. 2008. Effects of peripheral cooling on characteristics of local muscle. *Med Sport Sci* 53: 74-88.
- 16) Racinais S and Oksa J. 2010. Temperature and neuromuscular function. *Scand J Med Sci Sports* 20 Suppl 3: 1-18.
- 17) Clarke RS, Hellon RF and Lind AR. 1958. The duration of sustained contractions of the human forearm at different muscle temperatures. *J Physiol* 143: 454-473.

- 18) Bergh U and Ekblom B. 1979. Influence of muscle temperature on maximal muscle strength and power output in human skeletal muscles. *Acta Physiol Scand* 107: 33-37.
- 19) Davies CT, Mecrow IK and White MJ. 1982. Contractile properties of the human triceps surae with some observations on the effects of temperature and exercise. *Eur J Appl Physiol Occup Physiol* 49: 255-269.
- 20) Petrofsky JS and Lind AR. 1975. Insulative power of body fat on deep muscle temperatures and isometric endurance. *J Appl Physiol* 39: 639-642.
- 21) Sargeant AJ. 1987. Effect of muscle temperature on leg extension force and short-term power output in humans. *Eur J Appl Physiol Occup Physiol* 56: 693-698.
- 22) Oksa J, Rintamaki H and Rissanen S. 1997. Muscle performance and electromyogram activity of the lower leg muscles with different levels of cold exposure. *Eur J Appl Physiol Occup Physiol* 75: 484-490.
- 23) Oksa J, Rintamaki H, Makinen T, Hassi J and Rusko H. 1995. Cooling-induced changes in muscular performance and EMG activity of agonist and antagonist muscles. *Aviat Space Environ Med* 66: 26-31.
- 24) Blomstrand E, Bergh U, Essen-Gustavsson B and Ekblom B. 1984. Influence of low muscle temperature on muscle metabolism during intense dynamic exercise. *Acta Physiol Scand* 120: 229-236.
- 25) Blomstrand E and Essen-Gustavsson B. 1987. Influence of reduced muscle temperature on metabolism in type I and type II human muscle fibres during intensive exercise. *Acta Physiol Scand* 131: 569-574.
- 26) Bergh U and Ekblom B. 1979. Physical performance and peak aerobic power at different body temperatures. *J Appl Physiol Respir Environ Exerc Physiol* 46: 885-889.
- 27) Nadel ER, Holmer I, Bergh U, Astrand PO and Stolwijk JA. 1974. Energy exchanges of swimming man. *J Appl Physiol* 36: 465-471.
- 28) Holmer I and Bergh U. 1974. Metabolic and thermal response to swimming in water at varying temperatures. *J Appl Physiol* 37: 702-705.
- 29) De Jong RH, Hershey WN and Wagman IH. 1966. Nerve conduction velocity during hypothermia in man. *Anesthesiology* 27: 805-810.
- 30) Edwards RH, Harris RC, Hultman E, Kaijser L, Koh D and Nordesjo LO. 1972. Effect of temperature on muscle energy metabolism and endurance during successive isometric contractions, sustained to fatigue, of the quadriceps muscle in man. *J Physiol* 220: 335-352.
- 31) He ZH, Bottinelli R, Pellegrino MA, Ferenczi MA and Reggiani C. 2000. ATP consumption and efficiency of human single muscle fibers with different myosin isoform composition. *Biophys J* 79: 945-961.
- 32) Herve JC, Yamaoka K, Twist VW, Powell T, Ellory JC and Wang LC. 1992. Temperature dependence of electrophysiological properties of guinea pig and ground squirrel myocytes. *Am J Physiol* 263: R177-R184.
- 33) Kossler F, Lange F and Kuchler G. 1987. Isometric twitch and tetanic contraction of frog skeletal muscles at temperatures between 0 to 30 degrees C. *Biomed Biochim Acta* 46: 809-813.

- 34) Petrofsky JS and Lind AR. 1980. The influence of temperature on the amplitude and frequency components of the EMG during brief and sustained isometric contractions. *Eur J Appl Physiol Occup Physiol* 44: 189-200.
- 35) Petrofsky J and Laymon M. 2005. Muscle temperature and EMG amplitude and frequency during isometric exercise. *Aviat Space Environ Med* 76: 1024-1030.
- 36) Drinkwater EJ and Behm DG. 2007. Effects of 22 degrees C muscle temperature on voluntary and evoked muscle properties during and after high-intensity exercise. *Appl Physiol Nutr Metab* 32: 1043-1051.
- 37) Winkel J and Jorgensen K. 1991. Significance of skin temperature changes in surface electromyography. *Eur J Appl Physiol Occup Physiol* 63: 345-348.
- 38) Bell DG. 1993. The influence of air temperature on the EMG/force relationship of the quadriceps. *Eur J Appl Physiol Occup Physiol* 67: 256-260.
- 39) Mucke R and Heuer D. 1989. Behaviour of EMG-parameters and conduction velocity in contractions with different muscle temperatures. *Biomed Biochim Acta* 48: S459-S464.
- 40) Vincent MJ and Tipton MJ. 1988. The effects of cold immersion and hand protection on grip strength. *Aviat Space Environ Med* 59: 738-741.
- 41) Rome LC. 1990. Influence of temperature on muscle recruitment and muscle function in vivo. *Am J Physiol* 259: R210-R222.
- 42) Rome LC, Loughna PT and Goldspink G. 1984. Muscle fiber activity in carp as a function of swimming speed and muscle temperature. *Am J Physiol* 247: R272-R279.
- 43) Rome LC, Choi IH, Lutz G and Sosnicki A. 1992. The influence of temperature on muscle function in the fast swimming scup. I. Shortening velocity and muscle recruitment during swimming. *J Exp Biol* 163: 259-279.
- 44) Ranatunga KW. 1984. The force-velocity relation of rat fast and slow-twitch muscles examined at different temperatures. *J Physiol* 351: 517-529.
- 45) Holewijn M and Heus R. 1992. Effects of temperature on electromyogram and muscle function. *Eur J Appl Physiol Occup Physiol* 65: 541-545.
- 46) Bigland-Ritchie B, Donovan EF and Roussos CS. 1981. Conduction velocity and EMG power spectrum changes in fatigue of sustained maximal efforts. *J Appl Physiol Respir Environ Exerc Physiol* 51: 1300-1305.
- 47) Marsden CD, Meadows JC and Merton PA. 1983. "Muscular wisdom" that minimizes fatigue during prolonged effort in man: peak rates of motoneuron discharge and slowing of discharge during fatigue. *Adv Neurol* 39: 169-211.
- 48) Bawa P, Matthews PBC and Mekjavic IB. 1987. Electromyographic activity during shivering of muscles acting at the human elbow. *J Therm Biol* 12: 1-4.

- 49) Meigal AY, Oksa J, Hohtola E, Lupandin YV and Rintamaki H. 1998. Influence of cold shivering on fine motor control in the upper limb. *Acta Physiol Scand* 163: 41-47.
- 50) Pozos RS and Danzl D. 2001. Human physiological responses to cold stress and hypothermia. *Medical Aspects of Harsh Environments* 1: 351-382.
- 51) Hong SI and Nadel ER. 1979. Thermogenic control during exercise in a cold environment. *J Appl Physiol Respir Environ Exerc Physiol* 47: 1084-1089.
- 52) Abramson DI, Kahn A, Tuck S Jr, Turman GA, Rejal H and Fleischer CJ. 1958. Relationship between a range of tissue temperature and local oxygen uptake in the human forearm. I. Changes observed under resting conditions. *J Clin Invest* 37: 1031-1038.
- 53) Thorsson O, Lilja B, Ahlgren L, Hemdal B and Westlin N. 1985. The effect of local cold application on intramuscular blood flow at rest and after running. *Med Sci Sports Exerc* 17: 710-713.
- 54) Gregson W, Black MA, Jones H, Milson J, Morton J, Dawson B, Atkinson G and Green DJ. 2011. Influence of cold water immersion on limb and cutaneous blood flow at rest. *Am J Sports Med* 39: 1316-1323.
- 55) Rennie D, Park Y, Veicsteinas A and Pendergast D. 1980. Metabolic and circulatory adaptation to cold water stress. In: *Exercise Bioenergetics and Gas Exchange* (Cerretelli P, Whipp BJ, eds.): 315-321, Elsevier/North-Holland Biomedical Press, Amsterdam.
- 56) Shiojiri T, Shibasaki M, Aoki K, Kondo N and Koga S. 1997. Effects of reduced muscle temperature on the oxygen uptake kinetics at the start of exercise. *Acta Physiol Scand* 159: 327-333.
- 57) Whipp BJ, Ward SA, Lamarra N, Davis JA and Wasserman K. 1982. Parameters of ventilatory and gas exchange dynamics during exercise. *J Appl Physiol Respir Environ Exerc Physiol* 52: 1506-1513.
- 58) Ishii M, Ferretti G and Cerretelli P. 1992. Effects of muscle temperature on the VO<sub>2</sub> kinetics at the onset of exercise in man. *Respir Physiol* 88: 343-353.
- 59) Koga S, Wust RC, Walsh B, Kindig CA, Rossiter HB and Hogan MC. 2013. Increasing temperature speeds intracellular PO<sub>2</sub> kinetics during contractions in single *Xenopus* skeletal muscle fibers. *Am J Physiol Regul Integr Comp Physiol* 304: R59-R66.
- 60) Binzoni T and Delpy D. 2001. Local temperature changes and human skeletal muscle metabolism. *J Physiol Anthropol Appl Human Sci* 20: 159-174.
- 61) Binzoni T, Ngo L, Hiltbrand E, Springett R and Delpy D. 2002. Non-standard O<sub>2</sub> consumption-temperature curves during rest and isometric exercise in human skeletal muscle. *Comp Biochem Physiol A* 132: 27-32.
- 62) Hom C, Vasquez P and Pozos RS. 2004. Peripheral skin temperature effects on muscle oxygen levels. *J Therm Biol* 29: 785-789.
- 63) Yanagisawa O, Homma T, Okuwaki T, Shimao D and Takahashi H. 2007. Effects of cooling on human skin and skeletal muscle. *Eur J Appl Physiol* 100: 737-745.

- 64) Yanagisawa O and Fukubayashi T. 2010. Diffusion-weighted magnetic resonance imaging reveals the effects of different cooling temperatures on the diffusion of water molecules and perfusion within human skeletal muscle. *Clin Radiol* 65: 874-880.
- 65) Yanagisawa O, Takahashi H and Fukubayashi T. 2010. Effects of different cooling treatments on water diffusion, microcirculation, and water content within exercised muscles: evaluation by magnetic resonance T2-weighted and diffusion-weighted imaging. *J Sports Sci* 28: 1157-1163.
- 66) Hammel HT. 1964. Terrestrial animals in cold: recent studies of primitive man. In: *Hand Book of Physiology* (Dill DB and Adolph EF, eds.): 413-434, American Physiological Society, Washington DC.
- 67) LeBlanc J. 1978. Adaptation of man to cold. In: *Strategies in cold* (Wang LCH and Hudson JW, eds.): 695-715, Academic, New York.
- 68) Bittel JH. 1987. Heat debt as an index for cold adaptation in men. *J Appl Physiol* 62: 1627-1634.
- 69) Tipton MJ, Pandolf KB, Sawka MN, Werner J and Taylor NA. 2008. Physiological adaptation to hot and cold environments. In: *Physiological bases of human performance during work and exercise* (Taylor NAS and Groeller H, eds.): 379-400, Churchill Livingstone Elsevier.
- 70) Leblanc J. 1988. Factors affecting cold acclimation and thermogenesis in man. *Med Sci Sports Exerc* 20: S193-S196.
- 71) Golden FS and Tipton MJ. 1988. Human adaptation to repeated cold immersions. *J Physiol* 396: 349-363.
- 72) Budd GM, Brotherhood JR, Beasley FA, Hendrie AL, Jeffery SE, Lincoln GJ and Solaga AT. 1993. Effects of acclimatization to cold baths on men's responses to whole-body cooling in air. *Eur J Appl Physiol Occup Physiol* 67: 438-449.
- 73) Tipton MJ, Wakabayashi H, Barwood MJ, Eglin CM, Mekjavic IB and Taylor NA. 2013. Habituation of the metabolic and ventilatory responses to cold-water immersion in humans. *J Therm Biol* 38: 24-31.
- 74) Mäkinen TM, Rintamäki H, Korpelainen JT, Kampman V, Pääkkönen T, Oksa J, Palinkas LA, Leppäluoto J and Hassi J. 2005. Postural sway during single and repeated cold exposures. *Aviat Space Environ Med* 76: 947-953.
- 75) Muller MD, Seo Y, Kim CH, Ryan EJ, Pollock BS, Burns KJ and Glickman EL. 2014. Cold habituation does not improve manual dexterity during rest and exercise in 5 degrees C. *Int J Biometeorol* 58: 383-394.
- 76) Westerlund T, Oksa J, Smolander J and Mikkelsen M. 2009. Neuromuscular adaptation after repeated exposure to wholebody cryotherapy (-110°C). *J Therm Biol* 34: 226-231.
- 77) Livingstone SD. 1976. Changes in cold-induced vasodilation during Arctic exercises. *J Appl Physiol* 40: 455-457.
- 78) O'Brien C, Young AJ, Lee DT, Shitzer A, Sawka MN and Pandolf KB. 2000. Role of core temperature as a stimulus for cold acclimation during repeated immersion in 20 degrees C water. *J Appl Physiol* 89: 242-250.

- 79) Geurts CL, Sleivert GG and Cheung SS. 2005. Local cold acclimation of the hand impairs thermal responses of the finger without improving hand neuromuscular function. *Acta Physiol Scand* 183: 117-124.
- 80) Wakabayashi H, Wijayanto T, Kuroki H, Lee JY and Tochiyama Y. 2012. The effect of repeated mild cold water immersions on the adaptation of the vasomotor responses. *Int J Biometeorol* 56: 631-637.
- 81) Geurts CL, Sleivert GG and Cheung SS. 2006. Local cold acclimation during exercise and its effect on neuromuscular function of the hand. *Appl Physiol Nutr Metab* 31: 717-725.
- 82) Adams T and Smith RE. 1962. Effect of chronic local cold exposure on finger temperature responses. *J Appl Physiol* 17: 317-322.
- 83) Brajkovic D and Ducharme MB. 2003. Finger dexterity, skin temperature, and blood flow during auxiliary heating in the cold. *J Appl Physiol* 95: 758-770.
- 84) Ducharme MB, Brajkovic D and Frim J. 1999. The effect of direct and indirect hand heating on finger blood flow and dexterity during cold exposure. *J Therm Biol* 24: 391-396.
- 85) Geurts CL, Sleivert GG and Cheung SS. 2006. Central and peripheral factors in thermal, neuromuscular, and perceptual adaptation of the hand to repeated cold exposures. *Appl Physiol Nutr Metab* 31: 110-117.
- 86) Savourey G, Clerc L, Vallerand AL, Leftheriotis G, Mehier H and Bittel JH. 1992. Blood flow and muscle bio-energetics by <sup>31</sup>P-nuclear magnetic resonance after local cold acclimation. *Eur J Appl Physiol Occup Physiol* 64: 127-133.
- 87) Muller MD, Kim CH, Bellar DM, Ryan EJ, Seo Y, Muller SM and Glickman EL. 2012. Effect of cold acclimatization on exercise economy in the cold. *Eur J Appl Physiol* 112: 795-800.
- 88) Bae KA, An NY, Kwon YW, Kim C, Yoon CS, Park SC and Kim CK. 2003. Muscle fibre size and capillarity in Korean diving women. *Acta Physiol Scand* 179: 167-172.
- 89) Walters TJ and Constable SH. 1993. Intermittent cold exposure causes a muscle-specific shift in the fiber type composition in rats. *J Appl Physiol* 75: 264-267.
- 90) Soni A and Katoch SS. 1997. Structural and metabolic changes in skeletal muscle of cold acclimated rats. *J Therm Biol* 22: 95-107.
- 91) Suzuki J, Gao M, Ohinata H, Kuroshima A and Koyama T. 1997. Chronic Cold Exposure Stimulates Microvascular Remodeling Preferentially in Oxidative Muscles in Rats. *Jpn J Physiol* 47: 513-520.
- 92) Deveci D and Egginton S. 2002. Differing mechanisms of cold-induced changes in capillary supply in m. tibialis anterior of rats and hamsters. *J Exp Biol* 205: 829-840.
- 93) Sillau AH, Aquin L, Lechner AJ, Bui MV and Banchemo N. 1980. Increased capillary supply in skeletal muscle of guinea pigs acclimated to cold. *Respir Physiol* 42: 233-245.
- 94) Deveci D and Egginton S. 2003. Cold exposure differentially stimulates angiogenesis in glycolytic and oxidative muscles of rats and hamsters. *Exp Physiol* 88: 741-746.

- 95) van Marken Lichtenbelt WD and Daanen HA. 2003. Cold-induced metabolism. *Curr Opin Clin Nutr Metab Care* 6: 469-475.
- 96) van Marken Lichtenbelt WD and Schrauwen P. 2011. Implications of nonshivering thermogenesis for energy balance regulation in humans. *Am J Physiol Regul Integr Comp Physiol* 301: R285-R296.
- 97) van der Lans AA, Hoeks J, Brans B, Vijgen GH, Visser MG, Vosselman MJ, Hansen J, Jørgensen JA, Wu J, Mottaghy FM, Schrauwen P and van Marken Lichtenbelt WD. 2013. Cold acclimation recruits human brown fat and increases nonshivering thermogenesis. *J Clin Invest* 123: 3395-3403.
- 98) Yoneshiro T, Aita S, Matsushita M, Kayahara T, Kameya T, Kawai Y, Iwanaga T and Saito M. 2013. Recruited brown adipose tissue as an antiobesity agent in humans. *J Clin Invest* 123: 3404-3408.
- 99) Wijers SLJ, Schrauwen P, Saris WHM and van Marken Lichtenbelt WD. 2009. Human skeletal muscle mitochondrial uncoupling is associated with cold induced adaptive thermogenesis. *PLoS ONE* 3: e1777.
- 100) Wijers SL, Schrauwen P, van Baak MA, Saris WH and van Marken Lichtenbelt WD. 2011. Beta-adrenergic receptor blockade does not inhibit cold-induced thermogenesis in humans: possible involvement of brown adipose tissue. *J Clin Endocrinol Metab* 96: E598-E605.