

1           **Infrared cameras overestimate skin temperature during**  
2                                   **rewarming from cold exposure**

3   **Matthew J Maley<sup>1,2\*</sup>, Andrew P Hunt<sup>3</sup>, Aaron J E Bach<sup>4</sup>, Clare M Eglin<sup>2</sup>, Joseph T**  
4   **Costello<sup>2</sup>**

5   <sup>1</sup>Department of Sport, Institute of Human Sciences, University of Wolverhampton,  
6   Walsall, UK. <sup>2</sup>School of Sport, Health and Exercise Science, University of Portsmouth,  
7   Portsmouth, UK. <sup>3</sup>Institute of Health and Biomedical Innovation, School of Exercise and  
8   Nutrition Sciences, Queensland University of Technology, Brisbane, Australia. <sup>4</sup>National  
9   Climate Change Adaptation Research Facility, Griffith University, Gold Coast, Australia.

10   **\*Corresponding author**

11   m.maley2@wlv.ac.uk

12

13 **Abstract**

14 **Objective**

15 The primary aim of this study was to assess the accuracy of an infrared camera and that  
16 of a skin thermistor, both commercially available. The study aimed to assess the  
17 agreement over a wide range of skin temperatures following cold exposure.

18 **Methods**

19 Fifty-two males placed their right hand in a thin plastic bag and immersed it in 8 °C water  
20 for 30 minutes whilst seated in an air temperature of 30 °C. Following hand immersion,  
21 participants removed the bag and rested their hand at heart level for ten minutes. Index  
22 finger skin temperature ( $T_{sk}$ ) was measured with a thermistor, affixed to the finger pad,  
23 and an infrared camera measured 1 cm distally to the thermistor. Agreement between the  
24 infrared camera and thermistor was assessed by mean difference (infrared camera minus  
25 thermistor) and 95 % limits of agreement analysis, accounting for the repeated measures  
26 over time. The clinically significant threshold for  $T_{sk}$  differences was set at  $\pm 0.5$  °C and  
27 limits of agreement  $\pm 1$  °C.

28 **Results**

29 As an average across all time points, the infrared camera recorded  $T_{sk}$  1.80 (SD 1.16) °C  
30 warmer than the thermistor, with 95 % limits of agreement ranging from -0.46 °C to 4.07  
31 °C.

32 **Conclusion**

33 Collectively, the results show the infrared camera overestimated  $T_{sk}$  at every time point  
34 following local cooling. Further, measurement of finger  $T_{sk}$  from the infrared camera  
35 consistently fell outside the acceptable level of agreement (i.e. mean difference  
36 exceeding  $\pm 0.5$  °C). Considering these results, infrared cameras may overestimate  
37 peripheral  $T_{sk}$  following cold exposure and clinicians and practitioners should, therefore,  
38 adjust their risk/withdrawal criteria accordingly.

39

40 **Keywords**

41 Infrared thermography; temperature measurement; thermoregulation; cold exposure;  
42 instrument validity

43

## 44 **Introduction**

45 Measurement of skin temperature ( $T_{sk}$ ) is routinely conducted in research investigating  
46 human responses to environmental extremes. Measuring  $T_{sk}$  enables researchers to  
47 profile and calculate a wide variety of responses. Examples include mean body  
48 temperature (Hardy and Du Bois, 1938; Jay et al., 2007), body heat content (Burton,  
49 1935), and relatively newer calculations such as the adaptive physiological strain index  
50 (Buller et al., 2018; Hunt et al., 2019).

51 Measurement of  $T_{sk}$  may be conducted with a wide variety of devices that are broadly  
52 classified as conductive or infrared. Conductive devices, primarily thermistors, are often  
53 the preferred method for measuring  $T_{sk}$  (Bach et al., 2015b, 2015a; James et al., 2014).  
54 Most thermistor-based systems have a negative temperature coefficient. That is, their  
55 resistivity decreases with increasing temperature. In a thermistor-based system, a signal  
56 of 35 mV per °C is typical; nearly 1000 times greater than a thermocouple-based system  
57 (Bull, 2008). Thermistors are known for their long-term stability (Togawa, 1989),  
58 producing an error of ~0.1 °C (Bull, 2008). In contrast, infrared thermography is a non-  
59 contact technique, which transforms the energy radiated from objects in the infrared band  
60 into an electronic video signal that can be displayed on a computer and stored  
61 (Hildebrandt et al., 2010; Meola and Carlomagno, 2004).

62 Infrared cameras have added advantages over conductive devices. For example, they  
63 are able to capture (image or video) and store large quantities of data, meaning different  
64 areas of skin can be analysed retrospectively (Fernández-Cuevas et al., 2015; Moreira et  
65 al., 2017). In contrast, conductive devices are fixed to a specific skin site. Additionally,

66 most conductive devices are wired and, therefore, participants or patients are tethered to  
67 a data logger and their movement potentially restricted.

68 As a result of the improved accuracy of infrared devices over recent years, and despite  
69 the increased cost relative to conductive devices, their use has become more frequent in  
70 human physiology research. Example studies include local cold exposure (Brändström et  
71 al., 2008; Costello et al., 2012b, 2012a; Hope et al., 2014), identification of sporting  
72 injuries (Hildebrandt et al., 2010), testing individuals with non-freezing cold injuries (NFCI)  
73 (Ahle et al., 1990; Eglin et al., 2013) or Raynaud's phenomenon (Ring and Ammer, 2012;  
74 Shepherd et al., 2019), as well as during medical operations (Mercer et al., 2010).  
75 However, under controlled laboratory conditions during exercise and/or warm  
76 environmental conditions, thermal imaging technology to date has shown not to be  
77 acceptable; that is  $T_{sk}$  is under- or overestimated compared with a number of conductive  
78 devices (Bach et al., 2015a, 2015b; Buono et al., 2007; Fenemor et al., 2019; James et  
79 al., 2014).

80 Even though infrared cameras have been utilised to measure  $T_{sk}$  during cold exposure  
81 (Costello et al., 2012a; Hope et al., 2014; Maley et al., 2014), the accuracy of infrared  
82 cameras for measuring  $T_{sk} < 33\text{ }^{\circ}\text{C}$  is not well established. Several studies have compared  
83  $T_{sk}$  results between an infrared device and a conductive device during cold exposure;  
84 however, these studies are limited by sample size, the methodology of cold application  
85 and incomplete statistical analysis to properly assess the validity of infrared devices  
86 (Buono et al., 2007; Kelechi et al., 2011; Korukçu and Kilic, 2009).

87

88 If  $T_{sk}$  is cooled to  $<15\text{ }^{\circ}\text{C}$  for a prolonged period then the risk of peripheral cold injuries is  
89 significantly increased (Eglin et al., 2013; House et al., 2000; Maley et al., 2017, 2014;  
90 Thomas and Oakley, 2001). Considering this, the accuracy of infrared devices in  
91 measuring  $T_{sk}$  is paramount in order to not expose an individual to an increased risk of  
92 peripheral cold injuries. Given the need for valid monitoring of cooled  $T_{sk}$  across a wide  
93 range of sport, medical and occupational settings, this investigation set out to compare  
94 the agreement of  $T_{sk}$  between a conductive device with that of a non-contact infrared  
95 device during recovery from cold exposure.

96 **Methods**

97 This study was given ethical approval from the University of Portsmouth Science Faculty  
98 Ethics Committee and complied with standards set in The Declaration of Helsinki (2013).  
99 The participants were made aware of the purpose, procedures and risks of the study  
100 before giving their informed written consent. Fifty-two male participants volunteered in the  
101 study; their physical characteristics are as follows (mean [SD]): age 20 [2] years, height  
102 of 177.5 [7.7] cm, body mass of 75.5 [13.2] kg and hand length of 20.3 [1.2] cm.  
103 Participants' height and body mass were measured using a stadiometer (Leicester,  
104 Bodycare, UK) and digital weighing scales (Ohaus I-10, Ohaus Corporation, USA),  
105 respectively. Length of participants' right hand was measured using a segmometer  
106 (Segmometer 4, Rosscraft, Canada).

107 Participants entered the climate controlled chamber (mean [SD] dry-bulb: 30.3 [0.9] °C,  
108 wet-bulb: 22.9 [0.9] °C, wet-bulb globe temperature: 25.1 [0.9] °C) and rested in a semi-  
109 recumbent position for 25 minutes whilst being instrumented. During the 25 minute rest  
110 period, a skin thermistor (Type EUS-U, Grant Instruments, UK), connected to a data  
111 logger (Squirrel 2020, Grant Instruments, UK), was affixed onto the participant's second  
112 finger pad of the right hand using a small piece of breathable tape (Transpore Tape, 3M™,  
113 USA). Participants then placed their hand into a plastic bag, immersed it to the styloid  
114 process in a water bath maintained at 35.0 [0.2] °C for five minutes. Following this,  
115 participants removed their hand from the water, still within the plastic bag, and  
116 immediately placed it in a stirred water bath maintained at 8.1 [0.1] °C. After 30 minutes  
117 of cooling, participants removed their hand from the water bath and plastic bag to allow  
118 spontaneous rewarming for ten minutes. During this period, participants rested their hand

119 at the level of the heart. A calibrated infrared camera (A320G, FLIR Systems, UK) was  
120 positioned on a level tripod perpendicular to the participant's hand at a distance of one  
121 meter in line with published guidelines (Moreira et al., 2017). The camera was calibrated  
122 within 12 months of use. Data were recorded to dedicated software (ThermaCAM™  
123 Researcher, FLIR Systems, UK) to allow offline analysis. Emissivity (0.98) and distance  
124 (1 m) was set in object parameters within the software in line with published guidelines  
125 (Moreira et al., 2017).  $T_{sk}$  was measured immediately distal to the thermistor using the  
126 spot measurement tool.

127 The same thermistor was used for each participant and checked for accuracy before  
128 experimental use at eight water temperatures (5 °C to 40 °C, at increments of 5 °C). The  
129 thermistor was held at these temperatures in a precision water bath (Grant Instruments,  
130 UK) and compared to a UKAS calibrated precision digital thermometer (T600, Digitron  
131 Ltd, UK). Across the temperature range, the thermistor deviated by 0.11 (0.03) °C from  
132 the UKAS calibrated precision digital thermometer.

### 133 **Statistical Analysis**

134 The difference in  $T_{sk}$  following hand immersion was evaluated with a repeated-measures  
135 analysis of variance with Tukey's tests for multiple comparisons. Statistical significance  
136 was accepted at  $\alpha < 0.05$ . Agreement between the infrared camera and thermistor was  
137 assessed by the mean difference (infrared camera minus thermistor) and 95 % limits of  
138 agreement analysis, accounting for the repeated measures over time (Bland and Altman,  
139 2007). The clinically significant threshold for  $T_{sk}$  differences was set at 0.5 °C (Bach et  
140 al., 2015a; Marins et al., 2014; Niu et al., 2001; Selfe et al., 2008). Therefore, an



141 acceptable level of agreement would be a mean difference of  $<0.5$  °C and 95% limits of  
142 agreement of 1 °C.

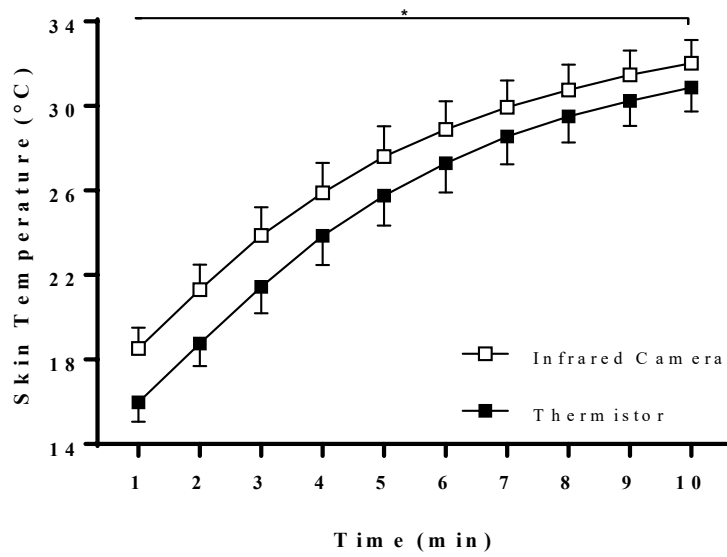
143 **Results**

144 A statistically significant interaction was observed between the measurement devices  
145 over time ( $F_{9,459} = 26.29$ ,  $P < 0.001$ ). Post-hoc analysis revealed the infrared camera was  
146 significantly higher than the thermistor at every minute during rewarming following hand  
147 immersion (Figure 1). As an average across all time points, the infrared camera recorded  
148  $T_{sk}$  1.80 (1.16) °C warmer than the thermistor, with 95 % limits of agreement ranging from  
149 -0.46 °C to 4.07 °C (Figure 2). At no time point was the acceptable level of agreement  
150 met (Table 1). At each minute, and as an average, mean differences were outside the  
151 acceptable level of agreement (Table 1).

152 Table 1. Mean difference and 95 % limits of agreement (LoA) for each time point and  
153 average

Time (min)	Mean Difference (°C)	Standard Deviation (°C)	Lower 95 % LoA (°C)	Upper 95 % LoA (°C)
1	2.55	1.32	-0.04	5.14
2	2.56	1.19	0.22	4.90
3	2.43	1.38	-0.27	5.14
4	2.04	1.12	-0.15	4.23
5	1.85	1.12	-0.35	4.04
6	1.61	0.99	-0.32	3.54
7	1.38	0.88	-0.34	3.11
8	1.24	0.65	-0.03	2.51
9	1.23	0.68	-0.10	2.57
10	1.15	0.66	-0.14	2.43
<b>Average</b>	1.80	1.16	-0.46	4.07

154 *Note: mean differences calculated as infrared camera minus thermistor.*



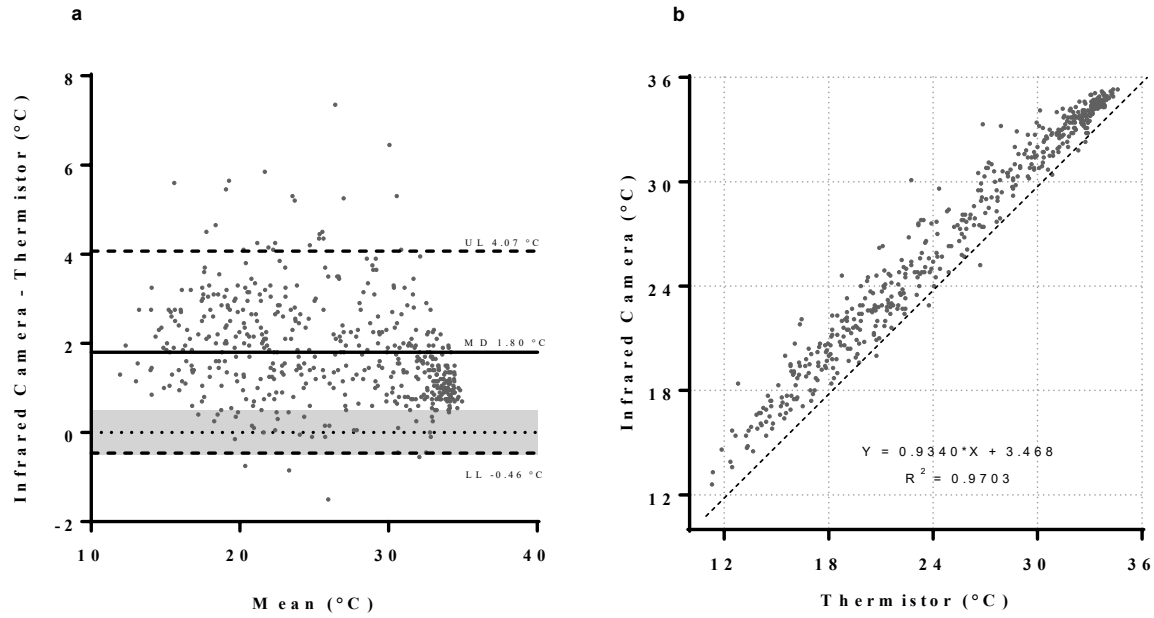
155

156 Figure 1. Mean (95 % confidence intervals) finger skin temperature measured by infrared  
 157 camera and thermistor following cold exposure

158 \*statistical difference between the infrared camera and thermistor ( $P < 0.001$ ).

159

160 Figure 2. Scatterplot (a) and Bland-Altman plot (b) of the agreement between the infrared



161 camera and thermistor for all time points

162 Note: MD, Mean difference; UL, Upper 95 % limits of agreement; LL, Lower 95 % limits of agreement. Grey  
163 band indicates a priori acceptable mean difference of 0.5 °C.

164

165 **Discussion**

166 The primary aim of this study was to assess the accuracy of an infrared camera with that  
167 of a skin thermistor, both of which are commercially available. The study aimed to assess  
168 the agreement over a wide range of  $T_{sk}$  following cold exposure; this was achieved as  $T_{sk}$   
169 ranged from 16 °C to 31 °C. Collectively, the results show the infrared camera  
170 overestimated  $T_{sk}$  at every time point following local cooling (Figure 1). Further,  
171 measurement of  $T_{sk}$  from the infrared camera consistently fell outside the acceptable level  
172 of agreement (i.e. mean difference  $>0.5$  °C) (Table 1, Figure 2b).

173 Few studies have previously assessed the accuracy of infrared devices with that of  
174 conductive devices during skin cooling. Buono et al. (2007) utilised a handheld infrared  
175 thermometer and contact thermistor to assess weighted mean  $T_{sk}$  during rest at air  
176 temperatures of 15 °C and 25 °C in six participants. They reported no statistical difference  
177 in  $T_{sk}$  between devices. Unfortunately, incomplete statistical analysis was conducted,  
178 preventing any meaningful interpretation of agreement between devices. Korukçu *et al.*  
179 (2009) tested facial  $T_{sk}$  of three participants using an infrared camera and contact  
180 thermocouple during mild car cabin cooling. Similar to Buono *et al.* (Buono et al., 2007),  
181 small sample size and inadequate statistical analysis were conducted preventing proper  
182 interpretation of agreement between devices, with authors summarising results of a  $<2$   
183 °C difference between the two devices. Finally, Kelechi *et al.* (2011) recruited 17  
184 participants and compared a handheld infrared device with a thermistor during local skin  
185 cooling of the legs. Following cooling, most measurements (71 %) had a mean difference  
186 of  $>0.5$  °C between devices. However, raw  $T_{sk}$  values are not provided which, similar to  
187 previous studies, makes interpretation of results difficult.

188 To the authors' knowledge, this is the first study to assess the agreement between  
189 infrared and conductive devices over a wide range of cool  $T_{sk}$  during a dynamic situation  
190 using a large sample. In this study, the infrared camera overestimated  $T_{sk}$  compared with  
191 the contact thermistor, which may have safety implications. As aforementioned, infrared  
192 devices have been used to measure  $T_{sk}$  following cryotherapy (Costello et al., 2012a,  
193 2012b; Selfe et al., 2014) and assess injury severity in NFCI patients (Ahle et al., 1990;  
194 Eglin et al., 2013; Thomas and Oakley, 2001). Cooling  $T_{sk}$  to  $<15\text{ }^{\circ}\text{C}$  for a prolonged  
195 period exposes an individual to local cold injuries, such as NFCI (Eglin et al., 2013; House  
196 et al., 2000; Maley et al., 2017, 2014; Thomas and Oakley, 2001). Thus, if a study uses  
197 an infrared camera to assess  $T_{sk}$  during or following cooling there is an increased risk of  
198 exposing that individual to lower than expected skin temperatures that may lead to NFCI.  
199 The authors are aware  $T_{sk}$  within this study did not reach  $<15\text{ }^{\circ}\text{C}$  but we can only speculate  
200 that the mean difference between devices would also be different at these lower skin  
201 temperatures.

202 The present study is not without limitations. Within the infrared camera software, the spot  
203 tool was chosen to record  $T_{sk}$ , which was distal to the contact thermistor location; meaning  
204 there was a difference in the location where  $T_{sk}$  was measured from. However, the authors  
205 are confident that the difference of around 1 cm between device measurement locations  
206 is not the reason for the overestimation of  $T_{sk}$  from the infrared camera (Maniar et al.,  
207 2015). The authors considered the possibility the contact thermistor was experiencing  
208 substantial thermal inertia, which may have explained why the thermistor consistently  
209 measured cooler  $T_{sk}$  compared with the infrared device. However, based on pilot studies,

210 the response rate of the thermistor ( $\Delta 15\text{ }^{\circ}\text{C}\cdot\text{min}^{-1}$ ) used far exceeds  $T_{sk}$  rewarming rates  
211 in this study.

## 212 **Conclusion**

213 In conclusion, the infrared camera utilised in this study overestimated  $T_{sk}$  between  $16\text{ }^{\circ}\text{C}$   
214 and  $31\text{ }^{\circ}\text{C}$ . Future research should consider using contact devices, checked for accuracy  
215 against a UKAS calibrated thermometer, in order to report accurate  $T_{sk}$  and to reduce the  
216 potential risk of peripheral cold injuries. Future research is needed to compare infrared  
217 and contact devices where  $T_{sk}$  is  $<15\text{ }^{\circ}\text{C}$ .

218

219 **Acknowledgements**

220 The authors thank the participants for their time and commitment to this study. The  
221 authors declare no conflicts of interest.

222 **Funding**

223 This research did not receive any specific grant from funding agencies in the public,  
224 commercial, or not-for-profit sectors.

225 **Ethical Statement**

226 This study was given ethical approval from the University of Portsmouth Science Faculty  
227 Ethics Committee and complied with standards set in The Declaration of Helsinki (2013).  
228 The participants were made aware of the purpose, procedures and risks of the study  
229 before giving their informed written consent.

230



231 **References**

- 232 Ahle, N.W., Buroni, J.R., Sharp, M.W., Hamlet, M.P., 1990. Infrared thermographic  
233 measurement of long term circulatory compromise in trenchfoot injured Argentine  
234 soldiers. *Aviat. Sp. Environ. Med.* 61, 247–250.
- 235 Bach, A.J.E., Stewart, I.B., Disher, A.E., Costello, J.T., 2015a. A comparison between  
236 conductive and infrared devices for measuring mean skin temperature at rest, during  
237 exercise in the heat, and recovery. *PLoS One* 10, 1–13.  
238 <https://doi.org/10.1371/journal.pone.0117907>
- 239 Bach, A.J.E., Stewart, I.B., Minett, G.M., Costello, J.T., 2015b. Does the technique  
240 employed for skin temperature assessment alter outcomes? A systematic review.  
241 *Physiol. Meas.* 36, R27–R51. <https://doi.org/10.1088/0967-3334/36/9/R27>
- 242 Bland, J.M., Altman, D.G., 2007. Agreement between methods of measurement with  
243 multiple observations per individual. *J. Biopharm. Stat.* 17, 571–582.  
244 <https://doi.org/10.1080/10543400701329422>
- 245 Brändström, H., Grip, H., Hallberg, P., Grönlund, C., Ångquist, K.-A., Giesbrecht, G.G.,  
246 2008. Hand cold recovery responses before and after 15 months of military training  
247 in a cold climate. *Aviat. Space. Environ. Med.* 79, 904–908.  
248 <https://doi.org/10.3357/ASEM.1886.2008>
- 249 Bull, K., 2008. Thermistors and thermocouples: matching the tool to the task in thermal  
250 validation. *J. Valid. Technol.* 14, 73–76.
- 251 Buller, M.J., Welles, A.P., Friedl, K.E., 2018. Wearable Physiological Monitoring for

252 Human Thermal-Work Strain Optimization. *J. Appl. Physiol.* 124, 432–441.  
253 <https://doi.org/10.1152/jappphysiol.00353.2017>

254 Buono, M.J., Jechort, A., Marques, R., Smith, C., Welch, J., 2007. Comparison of infrared  
255 versus contact thermometry for measuring skin temperature during exercise in the  
256 heat. *Physiol. Meas.* 28, 855–859. <https://doi.org/10.1088/0967-3334/28/8/008>

257 Burton, A.C., 1935. Human Calorimetry: II. The Average Temperature of the Tissues of  
258 the Body. *J. Nutr.* 9, 261–280.

259 Costello, J.T., Culligan, K., Selfe, J., Donnelly, A.E., 2012a. Muscle, Skin and Core  
260 Temperature after –110°C Cold Air and 8°C Water Treatment. *PLoS One* 7, e48190.

261 Costello, J.T., McInerney, C.D., Bleakley, C.M., Selfe, J., Donnelly, A.E., 2012b. The use  
262 of thermal imaging in assessing skin temperature following cryotherapy: a review. *J.*  
263 *Therm. Biol.* 37, 103–110. <https://doi.org/10.1016/j.jtherbio.2011.11.008>

264 Eglin, C.M., Golden, F.S.C., Tipton, M.J., 2013. Cold sensitivity test for individuals with  
265 non-freezing cold injury: the effect of prior exercise. *Extrem. Physiol. Med.* 2, 16.  
266 <https://doi.org/10.1186/2046-7648-2-16>

267 Fenemor, S.P., Gill, N.D., Sims, S.T., Beaven, C.M., Driller, M.W., 2019. Validity of a  
268 Tympanic Thermometer and Thermal Imaging Camera for Measuring Core and Skin  
269 Temperature during Exercise in the Heat. *Meas. Phys. Educ. Exerc. Sci.* 1–7.  
270 <https://doi.org/10.1080/1091367X.2019.1667361>

271 Fernández-Cuevas, I., Bouzas Marins, J.C., Arnáiz Lastras, J., Gómez Carmona, P.M.,

272 Piñonosa Cano, S., García-Concepción, M.Á., Sillero-Quintana, M., 2015.  
273 Classification of factors influencing the use of infrared thermography in humans: A  
274 review. *Infrared Phys. Technol.* <https://doi.org/10.1016/j.infrared.2015.02.007>

275 Hardy, J.D., Du Bois, E.F., 1938. The technic of measuring radiation and convection. *J.*  
276 *Nutr.* 15, 461–475.

277 Hildebrandt, C., Raschner, C., Ammer, K., 2010. An overview of recent application of  
278 medical infrared thermography in sports medicine in Austria. *Sensors* 10, 4700–  
279 4715. <https://doi.org/10.3390/s100504700>

280 Hope, K., Eglin, C.M., Golden, F., Tipton, M., 2014. Sublingual glyceryl trinitrate and the  
281 peripheral thermal responses in normal and cold-sensitive individuals. *Microvasc.*  
282 *Res.* 91, 84–89. <https://doi.org/10.1016/j.mvr.2013.11.002>

283 House, C.M., House, J.R., Oakley, E.H., 2000. Findings from a simulated disabled  
284 submarine survival trial. *Undersea Hyperb. Med.* 27, 175–183.

285 Hunt, A.P., Stewart, I.B., Billing, D.C., 2019. Indices of physiological strain for firefighters  
286 of the Australian Defence Forces. *J. Occup. Environ. Hyg.* 16, 727–734.  
287 <https://doi.org/10.1080/15459624.2019.1666211>

288 James, C.A., Richardson, A.J., Watt, P.W., Maxwell, N.S., 2014. Reliability and validity of  
289 skin temperature measurement by telemetry thermistors and a thermal camera  
290 during exercise in the heat. *J. Therm. Biol.* 45, 141–149.  
291 <https://doi.org/10.1016/j.jtherbio.2014.08.010>

292 Jay, O., Reardon, F.D., Webb, P., Ducharme, M.B., Ramsay, T., Nettlefold, L., Kenny,  
293 G.P., 2007. Estimating changes in mean body temperature for humans during  
294 exercise using core and skin temperatures is inaccurate even with a correction factor.  
295 J. Appl. Physiol. 103, 443–451. <https://doi.org/10.1152/jappphysiol.00117.2007>

296 Kelechi, T.J., Good, A., Mueller, M., 2011. Agreement and repeatability of an infrared  
297 thermometer. J. Nurs. Meas. 19, 55–64. <https://doi.org/10.1891/1061-3749.19.1.55>

298 Korukçu, M.Ö., Kilic, M., 2009. The usage of IR thermography for the temperature  
299 measurements inside an automobile cabin. Int. Commun. Heat Mass Transf. 36,  
300 872–877. <https://doi.org/https://doi.org/10.1016/j.icheatmasstransfer.2009.04.010>

301 Maley, M.J., Eglin, C.M., House, J.R., Tipton, M.J., 2014. The effect of ethnicity on the  
302 vascular responses to cold exposure of the extremities. Eur. J. Appl. Physiol. 114,  
303 2369–2379. <https://doi.org/10.1007/s00421-014-2962-2>

304 Maley, M.J., House, J.R., Tipton, M.J., Eglin, C.M., 2017. Role of cyclooxygenase in the  
305 vascular responses to extremity cooling in Caucasian and African males. Exp.  
306 Physiol. 102, 854–865. <https://doi.org/10.1113/EP086186>

307 Maniar, N., Bach, A.J.E., Stewart, I.B., Costello, J.T., 2015. The effect of using different  
308 regions of interest on local and mean skin temperature. J. Therm. Biol. 49–50, 33–  
309 38. <https://doi.org/10.1016/j.jtherbio.2015.01.008>

310 Marins, J.C.B., Fernandes, A.A., Cano, S.P., Moreira, D.G., da Silva, F.S., Costa, C.M.A.,  
311 Fernandez-Cuevas, I., Sillero-Quintana, M., 2014. Thermal body patterns for healthy  
312 Brazilian adults (male and female). J. Therm. Biol. 42, 1–8.

313 <https://doi.org/10.1016/j.jtherbio.2014.02.020>

314 Meola, C., Carlomagno, G.M., 2004. Recent advances in the use of infrared  
315 thermography. *Meas. Sci. Technol.* 15, R27–R58. [https://doi.org/10.1088/0957-](https://doi.org/10.1088/0957-0233/15/9/R01)  
316 [0233/15/9/R01](https://doi.org/10.1088/0957-0233/15/9/R01)

317 Mercer, J.B., Weerd, L. De, Miland, Å.O., Weum, S., 2010. Pre-, intra-, and postoperative  
318 use of dynamic infrared thermography (DIRT) provides valuable information on skin  
319 perfusion in perforator flaps used in reconstructive surgery, in: *Inframation*. Las  
320 Vegas, pp. 313–320.

321 Moreira, D.G., Costello, J.T., Brito, C.J., Adamczyk, J.G., Ammer, K., Bach, A.J.E., Costa,  
322 C.M.A., Eglin, C., Fernandes, A.A., Fernández-Cuevas, I., Ferreira, J.J.A., Formenti,  
323 D., Fournet, D., Havenith, G., Howell, K., Jung, A., Kenny, G.P., Kolosovas-Machuca,  
324 E.S., Maley, M.J., Merla, A., Pascoe, D., Priego-Quesada, J.I., Schwartz, R.G.,  
325 Seixas, A.R.D., Selfe, J., Vainer, B.G., Sillero-Quintana, M., 2017. Thermographic  
326 imaging in sports and exercise medicine: a Delphi study and consensus statement  
327 on the measurement of human skin temperature. *J. Therm. Biol.* 69, 155–162.  
328 <https://doi.org/10.1016/j.jtherbio.2017.07.006>

329 Niu, H.H., Lui, P.W., Hu, J.S., Ting, C.K., Yin, Y.C., Lo, Y.L., Liu, L., Lee, T.Y., 2001.  
330 Thermal symmetry of skin temperature: normative data of normal subjects in Taiwan.  
331 *Zhonghua Yi Xue Za Zhi (Taipei)*. 64, 459–468.

332 Ring, E.F.J., Ammer, K., 2012. Infrared thermal imaging in medicine. *Physiol. Meas.* 33,  
333 R33–R46. <https://doi.org/10.1088/0967-3334/33/3/R33>

334 Selfe, J., Alexander, J., Costello, J.T., May, K., Garratt, N., Atkins, S., Dillon, S., Hurst,  
335 H., Davison, M., Przybyla, D., Coley, A., Bitcon, M., Littler, G., Richards, J., 2014.  
336 The effect of three different (-135°C) whole body cryotherapy exposure durations on  
337 elite rugby league players. PLoS One 9, 1–9.  
338 <https://doi.org/10.1371/journal.pone.0086420>

339 Selfe, J., Whitaker, J., Hardaker, N., 2008. A narrative literature review identifying the  
340 minimum clinically important difference for skin temperature asymmetry at the knee.  
341 Thermol. Int. 18, 51–54.

342 Shepherd, A.I., Costello, J.T., Bailey, S.J., Bishop, N., Wadley, A.J., Young-Min, S.,  
343 Gilchrist, M., Mayes, H., White, D., Gorczynski, P., Saynor, Z.L., Massey, H., Eglin,  
344 C.M., 2019. “Beet” the cold: beetroot juice supplementation improves peripheral  
345 blood flow, endothelial function, and anti-inflammatory status in individuals with  
346 Raynaud’s phenomenon. J. Appl. Physiol. 127, 1478–1490.  
347 <https://doi.org/10.1152/jappphysiol.00292.2019>

348 Thomas, J.R., Oakley, H.N., 2001. Nonfreezing cold injury, in: Pandolf, K.B., Burr, R.E.  
349 (Eds.), Medical Aspects of Harsh Environments. Volume 1. TMM Publications,  
350 Washington, D.C., pp. 467–490.

351 Togawa, T., 1989. Non-contact skin emissivity: measurement from reflectance using step  
352 change in ambient radiation temperature. Clin. Phys. Physiol. Meas. 10, 39–48.

353