

1 **TITLE**

2 **Whole-body cryotherapy (-110°C) following high-intensity intermittent**
3 **exercise does not alter inflammatory, hormonal or muscle damage**
4 **biomarkers in trained males**

5

6 **RUNNING TITLE**

7 **Cryotherapy after exercise: Biomarkers of acute recovery**

8

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31 **ABSTRACT**

32 *Purpose:* This study examined the acute effects of a single session of
33 Whole-body Cryotherapy (WBC) following severe intermittent running
34 exercise on biomarkers of inflammation, muscle damage and stress.

35 *Methods:* Endurance-trained males (n=11) were tested twice using a within-
36 participant, balanced cross-over design that consisted of 5 x 5 min of high-
37 intensity running (HIR) followed by either 3 min of WBC at -110°C or a
38 passive control condition (CON). Before the HIR and after 60 min of recovery
39 a ramp-test was completed. At seven time points up to 24 hrs post exercise
40 venous blood samples were analyzed for serum levels of interleukin 6 (IL-
41 6), interleukin 10 (IL-10), c-reactive protein (CRP), soluble intercellular
42 adhesion molecule-1 (sICAM-1), myoglobin, cortisol, and testosterone.

43 *Results:* HIR induced significant increases in all biomarkers except sICAM-
44 1 in both recovery conditions, respectively. Compared to the CON condition
45 WBC did not attenuate exercise- induced changes in IL-6, IL-10, sICAM-1,
46 myoglobin, cortisol, testosterone or their ratio. Increased levels of cortisol
47 following exercise were negatively correlated with subsequent running
48 performance in both conditions (WBC: $r = -0.61$, $p = 0.04$; CON: $r = -0.64$, p
49 $= 0.04$).

50 *Conclusion:* The results of this study suggest that the postulated
51 physiological mechanisms by which WBC is proposed to improve recovery,
52 i.e. reductions in inflammation and muscle damage, may not be accurate.

53

54 **KEYWORDS**

55 Cryostimulation, Biomarkers, Cytokines, Acute recovery, Athletes

56

57 **HIGHLIGHTS**

58 • WBC did not affect changes in IL-6, IL-10, or myoglobin after high-

59 intensity exercise

60 • Similar data were recorded for testosterone, cortisol, and their ratio

61 • sICAM-1 was not altered by intermittent exercise or WBC

62 • Δ cortisol following exercise was negatively correlated with subsequent

63 performance

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65

66 **1. INTRODUCTION**

67 Whole-body Cryotherapy (WBC) or cryostimulation is a popular and widely
68 used recovery modality in sport and exercise medicine following intensive
69 training and competition. It consists of brief exposures (typically 2 to 4
70 minutes) to very cold air (-110°C and below) in cryogenic chambers with
71 individuals minimally dressed [1,2]. Originally utilized in a clinical setting for
72 treating symptoms of various rheumatic diseases, WBC is purported to
73 reduce pain, edema, and inflammation [3]. Therefore, WBC has become
74 very popular with both recreational and elite athletes.. To date, there is a
75 limited body of evidence regarding its efficacy and empirical data detailing
76 the potential mechanism(s) by which this treatment could be effective is
77 sparse [1].

78 Several authors have speculated that reductions in inflammation is the
79 primary mechanism by which WBC after strenuous exercise is believed to
80 be effective [1,2,5]. It is well established that intense exercise, especially if
81 the athlete is unaccustomed to such modalities and/or the exercise is
82 eccentric [6], leads to sarcomere disruptions and cell membrane damage
83 [7]. Following cell damage leukocytes are mobilized to the injured tissue by
84 soluble intercellular adhesion molecule-1 (sICAM-1), producing reactive
85 oxygen species and pro-inflammatory cytokines in the injured tissue,
86 resulting in intramuscular degradation and an amplification of muscle
87 damage [8]. This mechanism is defined as secondary muscle damage [8]
88 and may also be related to the increased levels of pro- and anti-

89 inflammatory cytokines observed following exercise [9]. It is believed that
90 exercise- induced inflammation, e.g. indicated by augmented Interleukin
91 (IL)-6 impairs athletic performance [10,11]. As WBC reduces skin-, muscle-
92 and core-temperature [12], leading to vasoconstriction and reduced blood
93 vessel permeability to immune cells, it is plausible that fewer leukocytes are
94 mobilized to the injured tissue, leading to a reduced pro-inflammatory
95 response and consequently less secondary muscle damage [13]. However,
96 the recent debate regarding the effects of WBC on modulating the
97 expression of sICAM-1 is still inconsistent and conflicting [5,14], possibly due
98 to the timing of the intervention post exercise. [14]To the best of our
99 knowledge, the effect of WBC on sICAM-1 following intense exercise has
100 not been compared to a control intervention.

101 Several studies have investigated the physical, psychological, and
102 physiological effects of WBC following exercise (for a review see [1,2,15])
103 Many [13,16–19], but not all [20,21] have reported that WBC might facilitate
104 the recovery process after exercise. Actually, for most biomarkers (e.g. pro-
105 and anti-inflammatory cytokines, creatin kinase (CK)) contradictory findings
106 have been reported in the literature. These conflicting results may be due to
107 large differences in methodology, such as exercise duration and intensity,
108 numbers of exercise bouts and WBC sessions and time points of biomarker
109 assessment. Thus, practical applications and recommendations for athletes
110 and their coaches are often difficult to conclude. As daily high-quality
111 performance and multiple competitions per week are required in many
112 sports, athletes often require interventions to enhance recovery within hours

113 or a few days. To date, only a few studies [13,16,19–23] investigated the
114 effects of a single WBC-treatment on acute recovery after high-intensity
115 exercise. In nine well-trained runners, performing a simulated trail run peak
116 torques of the knee extensors along with perceived sensation of pain and
117 tiredness was not significantly different in the WBC recovery condition
118 compared to passive rest [13]. With regard to biomarkers, no strong
119 conclusions can be made as alterations in some biomarkers (C-reactive
120 protein (CRP), Interleukin (IL)-1 β , IL-1 ra) indicated reduced inflammation,
121 while others (IL-6, IL-10, tumor necrosis factor (TNF)- α) remained
122 unchanged compared to passive recovery [19]. WBC applied after repeated
123 sprint exercise in professional soccer players induced a greater salivary
124 testosterone response compared to a control condition, but the changes did
125 not result in improvements in jump performance, blood lactate, CK
126 concentrations or perceived recovery [20]. [16][16]Other studies
127 demonstrated no beneficial effects of WBC on muscle force recovery [21] or
128 mixed results regarding perceived pain sensation and maximal physical
129 performance after hamstring damaging exercise [23]. As the majority of
130 previous studies focused on performance parameters it remains unclear
131 whether one session in combination with high-intensity exercise alters
132 biomarkers of hormonal status, inflammation and muscle damage.

133 Accordingly, the aim of the present study was to investigate the acute effects
134 of a single WBC-session during intermittent exercise on biomarkers
135 associated with exercise-induced inflammation, muscle damage and stress.
136 We hypothesized that a single exposure of WBC would reduce markers of

137 inflammation and muscle damage, and alter the cortisol-testosterone ratio
138 following high-intensity intermittent exercise

139

140 **2. METHODS**

141 **2.1 Participants**

142 A convenient sample size of 11 healthy, endurance-trained, male athletes
143 participated in the study (mean \pm SD age: 25.9 ± 2.1 yrs; height: 183.4 ± 3.4
144 cm; mass: 76.3 ± 6.6 kg; body mass index: 22.7 ± 1.7 kg·m⁻²; body fat: 10.7
145 ± 1.9 %; lean body mass: 68.1 ± 5.9 kg; peak oxygen uptake: 59.3 ± 5.3
146 mL·kg⁻¹·min⁻¹; performance level 3 and 4, according to De Pauw et al. [24]).
147 Inclusion criteria involved a history in endurance training (running) of at least
148 8 years with 3 or more training sessions per week as well as being familiar
149 with high-intensity interval training, i.e. short intervals of 2-8 min at 90-95%
150 of maximal heart rate, separated by equally short periods of recovery [25].
151 Athletes were excluded if they had any contraindications to WBC, such as
152 claustrophobia, cold hypersensitivity or abrasion injuries during medical
153 checkup as described elsewhere [2]. Participants were instructed to refrain
154 from consuming alcohol and caffeine 24 hrs prior to the tests and to maintain
155 their normal diet during the testing period. Intensive exercise was not
156 permitted for up to 48 hrs prior to the two test sessions and the individual
157 training was identical in the preceding week, respectively. Additional
158 recovery methods, including the use of non-steroidal anti-inflammatory
159 drugs, were not permitted. After demonstration and briefing about the

160 potential risks, all participants provided their written informed consent. The
161 study was conducted in accordance with the Declaration of Helsinki and
162 approved by the ethical committee of the German Sports University
163 Cologne.

164 **2.2 Experimental design**

165 This study was part of a larger project investigating effects of WBC on
166 performance variables, and the experimental design has been described in
167 detail elsewhere [16]. Briefly, a within-participant, balanced cross-over
168 design (with 7-days washout) was employed. Before participation, the
169 athletes underwent a medical checkup and familiarization with WBC.
170 Furthermore they carried out an incremental step test (starting velocity: 2.4
171 $\text{m}\cdot\text{s}^{-1}$; increase $0.4 \text{ m}\cdot\text{s}^{-1}$ every 5 min; treadmill gradient 1%) on a treadmill
172 (Woodway ELG 90/200 Sport, Lörrach, Germany) until individual exhaustion
173 to determine VO_2max (MetaLyzer 3b, Cortex, Leipzig, Germany) and the
174 individual intensities for High-intensity-running (HIR) during the two main
175 tests. The experimental protocol of these tests is presented in Figure 1.

176 ***** Figure 1 near here*****

177 All athletes were tested at the same time of day on each occasion and were
178 randomly assigned to start with either the WBC or control intervention (CON)
179 using research randomizer (version 4.0, retrieved from
180 <http://www.randomizer.org/>). The participants arrived at the laboratory at
181 least one hour prior to the test for acclimatization. At first, an incremental
182 exercise protocol to individual exhaustion was performed (Ramp 1). The

183 protocol consisted of 3 submaximal 3-min steps at 3.2, 3.6 and 4.0 m·s⁻¹ with
184 a treadmill gradient of 1% and 30 s of rest after each step. Thereafter velocity
185 was increased to 4.4 m·s⁻¹ and remained constant while the treadmill
186 gradient was increased by 0.5% every 30 s until exhaustion. After 5 min of
187 recovery, HIR was carried out, consisting of 5 x 5 min at 90 % of maximum
188 velocity (V_{max}) reached during the step test, with 4 min of active recovery in
189 between the intervals (60 % of V_{max}). HIR was followed by 1 h of passive
190 recovery, which was identically structured in both conditions except for the
191 implementation of one 3-min session of WBC after 45 min of rest. During the
192 recovery period the athletes remained seated in the conditioned laboratory
193 (ambient laboratory temperature WBC: 21.7 ± 0.8°C vs. CON: 21.7 ± 1.0°C;
194 humidity WBC: 36.4 ± 7.7% vs. CON: 35.8 ± 8.3%) and consumed 0.5 L of
195 a standardized fluid intake (energy: 400 kcal consisting of 46.5g
196 carbohydrates, 15g protein, 17g fat) to avoid dehydration and to replenish
197 depleted glycogen stores [26]. WBC was performed in a temperature-
198 controlled cryochamber with an electrical cooling system (Zimmer
199 MedizinSysteme GmbH, Ulm, Germany). The chamber system consists of
200 three separate rooms with constant temperatures of -10, -60 and -110°C and
201 we employed a protocol similar to that previously described elsewhere
202 [2,16]. The participants traversed the first two chambers with -10°C and -
203 60°C quickly and remained slowly walking for 3 min within the room at -
204 110°C. During the control intervention athletes walked slowly within the
205 laboratory for 3 min (at 21.7 ± 0.8°C and 35.8 ± 8.3 % humidity). After a total
206 of 60 min of recovery, athletes performed a second incremental exercise

207 (Ramp 2) with the same design as the first one. Time to exhaustion (t_{lim}) was
208 defined as the time (sec.) from the beginning of the ramp test until the
209 athlete's volitional exhaustion [16].

210 **2.3 Data measurement**

211 Before and after Ramp 1 + HIR ($R1_{pre}$, $R1_{post}$) and Ramp 2 ($R2_{pre}$, $R2_{post}$) as
212 well as 1-, 4- and 24-hrs after finishing the exercise protocol 8.5 mL venous
213 blood samples was obtained (BD Vacutainer Blood Collection System,
214 Beckton Dickson & Company, Plymouth, UK) from the antecubital vein
215 following 10 min of seated rest (see Figure 1). After collection, the samples
216 were stored at 7 °C for ~30-min for deactivation of coagulation factors before
217 being centrifuged (1861 g for 10-min at 4°C, Rotixa 50; Hettich Zentrifugen,
218 Mühlheim, Germany). The serum was then aliquoted (Eppendorf type) at -
219 80°C until later analysis. In particular, we were interested in the inflammatory
220 markers IL-6, IL-10, CRP and sICAM-1; the hormonal biomarkers cortisol
221 and testosterone, and the muscle damage biomarker myoglobin. Serum
222 levels of cortisol ($ng \cdot mL^{-1}$), testosterone ($ng \cdot mL^{-1}$), IL-6 ($pg \cdot mL^{-1}$), IL-10
223 ($pg \cdot mL^{-1}$), sICAM-1 ($ng \cdot mL^{-1}$), CRP ($mg \cdot L^{-1}$), and myoglobin ($ng \cdot mL^{-1}$) were
224 determined using human enzyme-linked immunosorbent assay (ELISA) kits.
225 Manufacturer instructions were followed for each of the kits and repeated
226 freeze-thaw cycles of serum were avoided. Intra-assay coefficient of
227 variations for cortisol, testosterone, c-reactive protein and Myoglobin (ELISA
228 kits manufactured by DRG Instruments GmbH, Marburg, Germany) as well
229 as sICAM-1 and IL-10 high sensitive (R&D Systems Inc, Minneapolis, USA)

230 and IL-6 high sensitive (IBL International GmbH, Hamburg, Germany) was
231 3.2%, 3.3%, 4.2%, 3.9%, 5.0%, 9.3%, and 4.6%, respectively. Minimum
232 detectable serum concentrations were 2.5 ng·mL⁻¹ for cortisol, 0.083 ng·mL⁻¹
233 ¹ for testosterone, 0.03 pg·mL⁻¹ for IL-6, 0.09 pg·mL⁻¹ for IL-10, 0.096 ng·mL⁻¹
234 ¹ for sICAM-1, 0.1 mg·L⁻¹ for CRP, and 5.0 ng·mL⁻¹ for myoglobin.
235 Hematological blood analysis was performed on the day of data collection
236 for the assessment of white blood cell count ($1 \cdot 10^3 \cdot \mu\text{L}^{-1}$) using Sysmex KX-
237 21N (Sysmex Deutschland GmbH, Norderstedt, Germany).

238 **2.4 Statistical Analyses**

239 All statistical tests were carried out using the Statistica software package for
240 Windows[®] (version 13.0, StatSoft Inc., Tulsa, OK, U.S.A). The distribution of
241 data was assessed using descriptive methods (skewness, outliers, and
242 distribution plots) and inferential statistics (Shapiro–Wilk test). As all data
243 were normally distributed data are presented as means \pm standard
244 deviations (SD). A two way (treatment [WBC, Control] * time [R1_{pre}, R1_{post},
245 R2_{pre}, R2_{post}, 1h, 4h, 24h]) repeated-measures analysis of variance
246 (ANOVA) was applied to compare all biomarkers. If main effects or
247 interactions were identified, Bonferroni post-hoc analysis was applied where
248 appropriate. Statistical significance was accepted at $P < 0.05$. Person
249 product-moment correlations were used to detect relationships between
250 ramp tests performance decrements ($\Delta t_{lim} = t_{lim}R2 - t_{lim}R1$) and changes in
251 cortisol, testosterone, IL-6, IL-10, sICAM-1, CRP, myoglobin, and white
252 blood cell count from baseline (R1_{pre}) to R2_{pre} in both recovery conditions,
253 respectively.

254

255 **3. RESULTS**

256 The serum concentrations of all biomarkers (mean \pm SD) are detailed in
257 table 1. At baseline (R1_{pre}), similar results were recorded for all outcome
258 measures (all $p > 0.05$) and all values were within normal range for healthy
259 individuals.

260 **3.1 Inflammatory Markers**

261 Significant time effects ($p < 0.01$) were observed for IL-6 (Fig 2A) and IL-10
262 (Fig 2B). No significant intervention or interaction effects were detected for
263 IL-6 ($p = 0.23$ and $p = 0.51$) and IL-10 ($p = 0.53$ and $p = 0.78$), respectively.
264 Compared to baseline (WBC: 0.85 ± 0.56 ; CON: 0.75 ± 0.41) IL-6 ($\text{pg}\cdot\text{mL}^{-1}$)
265 was significantly higher at R1_{post} (WBC: 2.12 ± 0.99 ; CON: 1.93 ± 0.51), R2_{pre}
266 (WBC: 1.19 ± 0.51 ; CON: 1.18 ± 0.28), R2_{post} (WBC: 1.22 ± 0.56 ; CON: 1.10
267 ± 0.31) and 1h (WBC: 1.25 ± 0.47 ; CON: 1.19 ± 0.42) (all $p < 0.01$). IL-10
268 ($\text{pg}\cdot\text{mL}^{-1}$) was elevated at R1_{post} (WBC: 3.62 ± 2.03 ; CON: 3.27 ± 0.97 , $p <$
269 0.01) and R2_{pre} (WBC: 3.03 ± 1.39 ; CON: 2.68 ± 0.61 , $p < 0.01$) compared
270 to baseline (WBC: 1.49 ± 0.42 ; CON: 1.44 ± 0.39).

271 Despite an increase 24h after exercise ($p < 0.01$), CRP (Fig 2C) was not
272 altered following WBC (intervention: $p = 0.51$; interaction: $p = 0.94$).

273 sICAM-1 levels following the two treatments remained similar (Fig. 2D), with
274 no intervention ($p = 0.96$) or interaction ($p = 0.27$) effects observed. Despite

275 observing a significant main effect for time ($p < 0.01$), no post-hoc
276 differences (all $p > 0.05$) using the Bonferroni correction were detected.

277 A significant increase over time ($p < 0.01$), but no intervention ($p = 0.73$) or
278 interaction ($p = 0.79$) effect was observed in white blood cell count (Fig. 2E).

279 Compared to baseline (WBC: 5.1 ± 1.4 ; CON: 5.4 ± 1.4) white blood cell
280 count ($\cdot 10^9 \cdot L^{-1}$) was elevated (all $p < 0.01$) at R1_{post} (WBC: 6.4 ± 1.7 ; CON:
281 6.3 ± 1.8) R2_{pre} (WBC: 7.4 ± 1.9 ; CON: 7.0 ± 2.2), R2_{post} (WBC: 10.7 ± 2.5 ;
282 CON: 10.0 ± 2.3), 1h (WBC: 9.5 ± 2.4 ; CON: 9.3 ± 2.0) and 4h (WBC: $9.1 \pm$
283 2.1 ; CON: 8.7 ± 1.6).

284 **3.2 Muscle damage**

285 There was a significant increase in myoglobin [$ng \cdot mL^{-1}$] over time ($p < 0.01$;
286 Fig. 2F); however no significant intervention ($p = 0.36$) or interaction ($p =$
287 0.73) effects were observed. Compared to baseline (WBC: 39.5 ± 7.7 ; CON:
288 39.8 ± 10.0) myoglobin was elevated at R1_{post} (WBC: 81.7 ± 26.2 ; CON: 84.5
289 ± 27.0), R2_{pre} (WBC: 94.1 ± 35.3 ; CON: 113.6 ± 52.8), R2_{post} (WBC: $93.6 \pm$
290 33.4 ; CON: 115.9 ± 64.3), 1h (WBC: 124.9 ± 52.2 ; CON: 158.0 ± 108.3) and
291 4h (WBC: 95.9 ± 31.4 ; CON: 123.0 ± 95.8) (all $p < 0.01$).

292 ***** Figure 2 near here *****

293 **3.3 Hormonal response**

294 A significant main effect over time was also overserved for cortisol ($p < 0.01$;
295 Fig 3A), testosterone ($p < 0.01$; Fig 3B), and testosterone to cortisol ratio (p
296 < 0.01 ; Fig 3C). Specifically, compared to baseline (WBC: 158.7 ± 26.7 ;

297 CON: 167.0 ± 29.8) cortisol ($\text{ng}\cdot\text{mL}^{-1}$) was elevated (all $p < 0.01$) after the
298 first ramp test (WBC: 217.9 ± 48.6 ; CON: 224.0 ± 58.2), at 1h (WBC: 115.6
299 ± 29.3 ; CON: 130.5 ± 14.6), 4h (WBC: 48.7 ± 18.4 ; CON: 62.8 ± 29.1) and
300 24h (WBC: 87.1 ± 20.3 ; CON: 89.0 ± 26.2). Compared to baseline (WBC:
301 5.2 ± 1.4 ; CON: 5.4 ± 1.9) testosterone ($\text{ng}\cdot\text{mL}^{-1}$) was also elevated (all $p <$
302 0.01) after the first ramp test (WBC: 6.4 ± 2.2 ; CON: 7.0 ± 2.1), at 1h (WBC:
303 4.0 ± 1.1 ; CON: 4.2 ± 1.4) and 4h (WBC: 3.2 ± 1.2 ; CON: 3.8 ± 1.3). Analysis
304 of the testosterone to cortisol ratio indicated that, compared to baseline
305 (WBC: 0.34 ± 0.11 ; CON: 0.35 ± 0.19) values increased significantly at 4h
306 (WBC: 0.80 ± 0.52 ; CON: 0.81 ± 0.54 , $p < 0.01$) and 24h (WBC: 0.63 ± 0.21 ;
307 CON: 0.72 ± 0.47 , $p < 0.01$). Again, no significant intervention or intervention
308 * time interaction was overserved for cortisol ($p = 0.53$ and $p = 0.93$
309 respectively), testosterone ($p = 0.67$ and $p = 0.81$ respectively), and the
310 testosterone to cortisol ratio ($p = 0.84$ and $p = 0.98$ respectively).

311 ***** Figure 3 near here*****

312

313 **3.4 Correlation**

314 Performance decrements (Δt_{lim}) correlated significantly with $\Delta\text{cortisol}$ after
315 both the CON ($r = -0.64$, $p = 0.04$) and the WBC interventions ($r = -0.61$, $p =$
316 0.04 ; Fig. 4). No statistically significant correlations were detected for Δt_{lim}
317 and the change in any of the other biomarkers (all $p > 0.05$).

318 ***** Figure 4 near here*****

319

*** Table 1 near here***

320 4. DISCUSSION

321 The current study is the most thorough investigation of the acute effects (up
322 to 24 hrs) of a single WBC-session following intermittent high-intensity
323 exercise on hormonal, inflammatory and muscle damage biomarkers to
324 date. The main findings of this investigation are as follows: (1) contrary to
325 our initial hypothesis, compared to passive recovery one session of WBC
326 did not alter the exercise-induced inflammatory, muscle damage or
327 hormonal response to high-intensity running in trained athletes, (2) the
328 exercise-induced perturbations of all inflammatory, muscle damage, and
329 hormonal biomarkers, except CRP and testosterone to cortisol ratio,
330 returned to basal levels within 24 hrs, and (3) increased levels of cortisol,
331 induced by high-intensity exercise, were negatively correlated to subsequent
332 running performance. Collectively, these data suggest that the postulated
333 physiological mechanism(s), i.e. reductions in inflammation and muscle
334 damage, by which WBC is purported to enhance recovery from EIMD
335 following high-intensity running in trained male athletes may not be accurate.

336 Reducing the inflammatory response following exercise is one of the primary
337 reasons why WBC is applied as a recovery method [2]. In the present study,
338 similar inflammatory responses were observed following WBC and the
339 control intervention, with IL-6 and IL-10 peaking immediately after exercise
340 and returning to baseline after 1-4 hrs post-exercise (see Fig 2). Pournot and
341 colleagues [19] have previously reported comparable IL-6 and IL-10

342 reactions following a simulated trail run on a treadmill in 11 well-trained
343 distance runners. While exercise duration was longer in our investigation
344 (56.3 ± 1.6 min vs 48 min), we did not include downhill running, that is known
345 to induce severe structural muscle damage and inflammation due to high
346 muscular load during eccentric contractions [13]. Selfe and colleagues [27]
347 also reported similar IL-6 values to those of the present investigation and no
348 effects of WBC on IL-6 concentration applied 10-16 hrs after a rugby training.
349 Interestingly, another study reported reduced IL-6 values after 40-min of
350 cycling in professional volleyball players following one session of WBC
351 compared to no WBC and the authors suggested that WBC might initiate
352 protective effects [28]. As WBC preceded exercise, it is likely that these
353 methodological differences caused the differences in the cytokine profile
354 compared to the present study.

355 Despite a main effect over time, sICAM-1 was not altered following the WBC
356 intervention (see Fig. 2). Thus, assuming sICAM-1 plays a key-role in the
357 inflammation response [5,14], the present data suggest that a single session
358 of WBC is insufficient to reduce the exercise-induced inflammation. Banfi
359 and colleagues [29] have previously reported reduced levels of sICAM-1
360 after five daily WBC- sessions in rugby players, yet due to the absence of a
361 control group, it is difficult to delineate if WBC in isolation was responsible
362 for these findings [29]. In contrast, Both Dugué and Leppänen [30] and
363 Buemi and colleagues [31] reported increased levels of sICAM-1 after the
364 application of cold water immersion. There is a brevity of empirical data

365 investigating the effects of WBC on the sICAM-1, especially repeated
366 exposures, thus this topic warrants further investigation [5,14].

367 In comparison to the control condition, WBC did not alter CRP (see Fig. 2).
368 Similarly, no effect of WBC on CRP response was reported in rugby players
369 after five days with continuous training and daily WBC- treatments [32].
370 Contrary, Pournot and colleagues observed reduced CRP-levels 24 hrs after
371 one session of WBC (3 min at -110°C) compared to passive recovery in
372 equally trained participants [19]. Overall higher CRP-values compared to
373 our results are most likely caused by additional eccentric contractions during
374 downhill running. Furthermore, Pournot reported decreased levels in pro-
375 inflammatory cytokine IL-1 β and increased levels in anti-inflammatory
376 cytokine IL-1ra, suggesting, in contrast to our findings, that one session of
377 WBC reduces the inflammatory process [19].

378 CK is probably the most frequently analyzed biomarker to identify muscle
379 damage [33]. CK only leaks into the bloodstream when the sarcolemma is
380 damaged and is, therefore, another commonly used biomarker of muscle
381 damage [34]. However, the enzyme has a molecular mass of 84 kilodaltons
382 (kDa) and has to be transported by the lymphatic system [6]. Therefore, the
383 onset of CK-concentration in venous blood is delayed. - Depending on the
384 magnitude of muscle damage, CK peaks 24 to 96 hrs after exercise [35].
385 Therefore myoglobin, a rather small molecule (18 kDa) that is released
386 directly to the blood flow as a result of degradation of muscle proteins [34],
387 appears to be more feasible to detect the acute muscle damage effects of

388 strenuous exercise. Myoglobin typically peaks within the first hours after
389 severe exercise and is already decreasing at 24 hrs post exercise whereas
390 CK is still rising [36,37]. To the best of our knowledge, this is the first
391 investigation that has utilized myoglobin as marker for muscle damage after
392 WBC. Myoglobin peaked 1 hr after exercise and returned to baseline after
393 24 hrs with no differences between recovery modalities (see Fig 2).
394 Although this is the first study to assess myoglobin, others have reported
395 comparable results regarding CK [13,20], but also reduced levels of CK after
396 WBC [3,18,38–40] [13,20]. However, these studies have some limitations,
397 such as small sample size [38], the lack of a control group [40] or the high
398 probability that the results were influenced by the repeated bout effect
399 [18,39]. Therefore, the evidence for WBC to reduce muscle damage remains
400 very limited.

401 Testosterone, cortisol, and their ratio are often employed as biomarkers of
402 anabolic status, training responses/adaptations and motivation [41]. It is well
403 established that cortisol increases when an individual is exposed to
404 psychophysiological stress following activation of the hypothalamic-pituitary-
405 adrenal axis [42]. The physical exercise itself potentially increases cortisol
406 further, though the magnitude depends on, amongst other variables, the type
407 of exercise [43] and the environmental conditions [44] where it takes place.
408 In the present study we observed increased levels in cortisol at baseline and
409 exercise induced increments in cortisol and testosterone, respectively (see
410 Fig 3). Due to heterogeneous methodologies and conflicting results in the
411 current literature, the effects of WBC on hormone biomarkers is still unclear

412 [38,45–47]. However, to our knowledge there is only one study which has
413 investigated the effects of a single WBC-session, applied immediately after
414 exercise, on cortisol and testosterone [20]. In line with the findings of the
415 present study, Russell and colleagues [20] detected no WBC- related
416 changes in cortisol. Collectively, these findings suggest that a single WBC-
417 session does not alter the stress-related hormone cortisol. However, in
418 contrast to our findings Russell and colleagues [20] reported increased
419 testosterone levels in the WBC- group, suggesting that higher testosterone-
420 concentrations may facilitate recovery.[20]. Moreover, the authors speculate
421 that higher testosterone- values might indicate reduced inflammation as low
422 serum- testosterone concentrations are related to inflammation [20]. These
423 results and speculations cannot be confirmed by the findings of the present
424 investigation, as testosterone responded similarly after the WBC and CON-
425 interventions. The contrasting findings might be explained by the application
426 of different exercise protocols. Immediately after repeated sprint exercise,
427 Russell et al [20] detected no time effects in cortisol or testosterone, while
428 our data indicated significant elevations in both hormones, most likely due
429 to higher intensity of the exercise intervention. Therefore, it is plausible to
430 speculate that testosterone concentrations are only elevated by one
431 session of WBC if the preceding exercise itself did not induce a significant
432 hormonal response.

433 The present study also investigated the relationship between the
434 performance decrements after HIR (i.e. $\Delta t_{lim} R1 - R2$) and the changes in
435 biomarker-levels from baseline to the end of 60 min recovery period ($R2_{pre}$

436 – $R1_{pre}$). No correlations were found for all biomarkers except cortisol,
437 regardless of the recovery modality applied (see Fig. 4). These results
438 suggest that reductions in cortisol after exercise and recovery lead to higher
439 subsequent running-performance. Interestingly, despite no significant
440 intervention or interaction effects in inflammatory, muscle damage and
441 stress-related biomarkers, we observed improved running performance
442 (time to exhaustion) immediately after a single WBC-session [16]. It can be
443 speculated that these performance improvements are most likely attributed
444 to i) a placebo effect, ii) perceptual reductions in pain, iii) acute changes in
445 muscle oxygenation, iv) lower cardiovascular strain, or v) a combination of
446 these factors.

447

448 **5. LIMITATIONS**

449 Our study has limitations that warrant mention. Firstly, incorporating a range
450 of other inflammatory cytokines (e.g. TNF-alpha, IL-8 and IL-15), and
451 biomarkers of muscle damage (e.g. creatine kinase, lactate dehydrogenase)
452 and stress (e.g. epinephrine, alpha amylase) would have provided further
453 insight into the inflammatory, damage, and hormonal effects of a single WBC
454 exposure following exercise. Secondly, despite all biomarkers except CRP
455 and testosterone-cortisol ratio returning to basal levels a longer timeline of
456 analysis, possibly up to 96 hrs post exercise, would have offered additional
457 insights into the potential effects of WBC. Thirdly, although the present
458 investigation focused on inflammatory, hormonal and muscle damage

459 biomarkers, functional and performance measures up to 24 hrs would have
460 added additional practical information regarding the application of WBC.
461 Fourthly, despite employing a cross-over study design, it is possible that the
462 small sample-size increased the potential for type II error. Furthermore, a
463 cross-over study design might be influenced by the repeated bout effect.
464 However, due to the participants being very familiar with the exercise
465 protocol and the lack of any unaccustomed exercise or downhill running, i.e.
466 eccentric muscle damage, it is very unlikely that these results were impacted
467 by the repeated bout effect. Future research incorporating a parallel design
468 with a larger sample size is therefore warranted. Finally, not including an
469 active recovery group is a limitation of this study.

470

471 **6. CONCLUSION**

472 This study is the most thorough investigation of the effects of a single
473 session of WBC (3 min at -110°C) on biomarkers of hormonal status,
474 inflammation, and muscle damage after acute high-intensity exercise in
475 trained males. Despite the expected changes in IL-6, IL-10, CRP, sICAM-1,
476 myoglobin, cortisol, testosterone, and cortisol-testosterone ratio in the 24 hrs
477 following exercise, and contrary to our hypotheses, our results demonstrate
478 for the first time that WBC has no acute beneficial effect compared to passive
479 recovery in any biomarker assessed. The results of this study suggest that
480 the postulated physiological mechanisms by which a single exposure to

481 WBC is speculated to improve recovery, i.e. reductions in inflammation and
482 muscle damage, may not be accurate.

483

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487 granting access to their cryochamber and performance center.


488

489 **DISCLOSURE STATEMENT**

490 The authors confirm there are no conflicts of interest.

491

492 **Figure captions**

493 **Figure 1.** Schematic presentation of the experimental randomized cross
494 over design.  denotes blood sampling at seven time points, before
495 and after ramp test 1 (R1_{pre}; R1_{post}) and 2 (R2_{pre}; R2_{post}) and 1 (1h), 4 (4h)
496 and 24 (24h) hrs after the intervention for whole-body cryotherapy (WBC)
497 and control (CON) intervention, respectively.

498 **Figure 2.** Serum concentrations (mean \pm SD) of interleukin 6 (IL-6; A),
499 interleukin 10 (IL-10; B), C-reactive Protein (CRP; C), soluble intercellular
500 adhesion molecule-1 (sICAM-1; D) and myoglobin (F) as well as white blood
501 cell count (WBC-count; E) at seven time points (R1_{pre}; R1_{post}; R2_{pre}; R2_{post};
502 1h; 4h; 24h). * P < 0.05 time effect compared to baseline (R1_{pre}), for both

503 interventions (whole-body cryotherapy (WBC) and control (CON))
504 combined.

505 **Figure 3.** Serum concentrations (mean \pm SD) of cortisol (A), testosterone
506 (B) and calculation of testosterone to cortisol ratio (C) at seven time points
507 ($R1_{pre}$; $R1_{post}$; $R2_{pre}$; $R2_{post}$; 1h; 4h; 24h). * $P < 0.05$ time effect compared to
508 baseline ($R1_{pre}$), for both interventions (whole-body cryotherapy [WBC] and
509 control [CON]) combined.

510 **Figure 4.** Correlations between change (Δ) in time to exhaustion (t_{lim}) from
511 ramp 1 to ramp 2 and change in serum cortisol from $R1_{pre}$ to $R2_{pre}$ in control
512 (CON) and whole-body cryotherapy (WBC) intervention, respectively.

513

514 REFERENCES

515

- 516 [1] J.T. Costello, P.R.A. Baker, G.M. Minett, F. Bieuzen, I.B. Stewart, C.
517 Bleakley, Whole-body cryotherapy (extreme cold air exposure) for preventing
518 and treating muscle soreness after exercise in adults., Cochrane Database of
519 Systematic Reviews (9) (2015).
- 520 [2] G. Lombardi, E. Ziemann, G. Banfi, Whole-Body Cryotherapy in Athletes:
521 From Therapy to Stimulation. An Updated Review of the Literature, *Frontiers*
522 *in physiology* 8 (2017) 258.
- 523 [3] G. Banfi, G. Lombardi, A. Colombini, G. Melegati, Whole-body cryotherapy in
524 athletes, *Sports medicine (Auckland, N.Z.)* 40 (6) (2010) 509–517.
- 525 [4] W. Poppendieck, O. Faude, M. Wegmann, T. Meyer, Cooling and
526 performance recovery of trained athletes: a meta-analytical review,
527 *International journal of sports physiology and performance* 8 (3) (2013) 227–
528 242.
- 529 [5] J.B. Ferreira-Junior, M. Bottaro, J.P. Loenneke, A. Vieira, C.A. Vieira, M.G.
530 Bembem, Could whole-body cryotherapy (below -100°C) improve muscle
531 recovery from muscle damage?, *Frontiers in physiology* 5 (2014) 247.
- 532 [6] M. Behringer, J. Montag, Y. Kilian, M. Mccourt, J. Mester, The repeated bout
533 effect: Is the blunted creatine kinase response an effect of an altered enzyme

- 534 inactivation kinetic?, *The Journal of sports medicine and physical fitness* 55
535 (12) (2015) 1431–1437.
- 536 [7] A.L. Mackey, A.E. Donnelly, H.P. Roper, Muscle connective tissue content of
537 endurance-trained and inactive individuals, *Scandinavian journal of medicine*
538 & science in sports 15 (6) (2005) 402–408.
- 539 [8] G. Paulsen, U.R. Mikkelsen, T. Raastad, J.M. Peake, Leucocytes, cytokines
540 and satellite cells: What role do they play in muscle damage and
541 regeneration following eccentric exercise?, *Exercise immunology review* 18
542 (2012) 42–97.
- 543 [9] P.M. Clarkson, M.J. Hubal, Exercise-induced muscle damage in humans,
544 *American journal of physical medicine & rehabilitation* 81 (11 Suppl) (2002)
545 S52-69.
- 546 [10] N.T. Vargas, F. Marino, A neuroinflammatory model for acute fatigue during
547 exercise, *Sports medicine (Auckland, N.Z.)* 44 (11) (2014) 1479–1487.
- 548 [11] P.J. Robson-Ansley, L. de Milander, M. Collins, T.D. Noakes, Acute
549 interleukin-6 administration impairs athletic performance in healthy, trained
550 male runners, *Canadian journal of applied physiology* 29 (4) (2004) 411–418.
- 551 [12] J.T. Costello, K. Culligan, J. Selfe, A.E. Donnelly, Muscle, skin and core
552 temperature after -110°C cold air and 8°C water treatment, *PloS one* 7 (11)
553 (2012) e48190.
- 554 [13] C. Hausswirth, J. Louis, F. Bieuzen, H. Pournot, J. Fournier, J.-R. Filliard, J.
555 Brisswalter, Effects of whole-body cryotherapy vs. far-infrared vs. passive
556 modalities on recovery from exercise-induced muscle damage in highly-
557 trained runners, *PloS one* 6 (12) (2011) e27749.
- 558 [14] B.M. Dugué, An attempt to improve Ferreira-Junior model concerning the
559 anti-inflammatory action of whole-body cryotherapy after exercise induced
560 muscular damage (EIMD), *Frontiers in physiology* 6 (2015) 35.
- 561 [15] C.M. Bleakley, F. Bieuzen, G.W. Davison, J.T. Costello, Whole-body
562 cryotherapy: empirical evidence and theoretical perspectives, *Open access*
563 *journal of sports medicine* 5 (2014) 25–36.
- 564 [16] M. Kruger, M. de Marees, K.-H. Dittmar, B. Sperlich, J. Mester, Whole-body
565 cryotherapy's enhancement of acute recovery of running performance in well-
566 trained athletes, *International journal of sports physiology and performance*
567 10 (5) (2015) 605–612.
- 568 [17] J.B. Ferreira-Junior, M. Bottaro, A. Vieira, A.F. Siqueira, C.A. Vieira, Durigan,
569 J L Q, E.L. Cadore, Coelho, L G M, H.G. Simões, M.G. Bembem, One
570 session of partial-body cryotherapy (-110 °C) improves muscle damage
571 recovery, *Scandinavian journal of medicine & science in sports* (2014).
- 572 [18] E. Ziemann, R.A. Olek, T. Grzywacz, J.J. Kaczor, J. Antosiewicz, W. Skrobot,
573 S. Kujach, R. Laskowski, Whole-body cryostimulation as an effective way of
574 reducing exercise-induced inflammation and blood cholesterol in young men,
575 *European cytokine network* 25 (1) (2014) 14–23.
- 576 [19] H. Pournot, F. Bieuzen, J. Louis, R. Mounier, J.-R. Fillard, E. Barbiche, C.
577 Hausswirth, Time-course of changes in inflammatory response after whole-

- 578 body cryotherapy multi exposures following severe exercise, PloS one 6 (7)
579 (2011) e22748.
- 580 [20] M. Russell, J. Birch, T. Love, C.J. Cook, R.M. Bracken, T. Taylor, E. Swift, E.
581 Cockburn, C. Finn, D. Cunningham, L. Wilson, L.P. Kilduff, The Effects of a
582 Single Whole-Body Cryotherapy Exposure on Physiological, Performance,
583 and Perceptual Responses of Professional Academy Soccer Players After
584 Repeated Sprint Exercise, *Journal of strength and conditioning research* 31
585 (2) (2017) 415–421.
- 586 [21] J.T. Costello, L.A. Algar, A.E. Donnelly, Effects of whole-body cryotherapy (-
587 110 °C) on proprioception and indices of muscle damage, *Scandinavian
588 journal of medicine & science in sports* 22 (2) (2012) 190–198.
- 589 [22] J.B. Ferreira-Junior, M. Bottaro, C.A. Vieira, Soares, S R S, A. Vieira, V.A.
590 Cleto, E.L. Cadore, D.B. Coelho, H.G. Simoes, L.E. Brown, Effects of partial-
591 body cryotherapy (- 110°C) on muscle recovery between high-intensity
592 exercise bouts, *International journal of sports medicine* 35 (14) (2014) 1155–
593 1160.
- 594 [23] B. Fonda, N. Sarabon, Effects of whole-body cryotherapy on recovery after
595 hamstring damaging exercise: a crossover study, *Scandinavian journal of
596 medicine & science in sports* 23 (5) (2013) 8.
- 597 [24] K. de Pauw, B. Roelands, S.S. Cheung, B. de Geus, G. Rietjens, R.
598 Meeusen, Guidelines to classify subject groups in sport-science research,
599 *International journal of sports physiology and performance* 8 (2) (2013) 111–
600 122.
- 601 [25] M. Buchheit, P.B. Laursen, High-intensity interval training, solutions to the
602 programming puzzle: Part I: cardiopulmonary emphasis, *Sports medicine
603 (Auckland, N.Z.)* 43 (5) (2013) 313–338.
- 604 [26] K.J. Spaccarotella, W.D. Andzel, The effects of low fat chocolate milk on
605 postexercise recovery in collegiate athletes, *Journal of strength and
606 conditioning research* 25 (12) (2011) 3456–3460.
- 607 [27] J. Selfe, J. Alexander, J.T. Costello, K. May, N. Garratt, S. Atkins, S. Dillon,
608 H. Hurst, M. Davison, D. Przybyla, A. Coley, M. Bitcon, G. Littler, J. Richards,
609 The effect of three different (-135°C) whole body cryotherapy exposure
610 durations on elite rugby league players, *PloS one* 9 (1) (2014) e86420.
- 611 [28] C. Mila-Kierzenkowska, A. Jurecka, A. Woźniak, M. Szpinda, B.
612 Augustyńska, B. Woźniak, The effect of submaximal exercise preceded by
613 single whole-body cryotherapy on the markers of oxidative stress and
614 inflammation in blood of volleyball players, *Oxidative medicine and cellular
615 longevity* 2013 (2013) 409567.
- 616 [29] G. Banfi, G. Melegati, A. Barassi, G. Dogliotti, G. Melzi d’Eril, B. Dugué, M.M.
617 Corsi, Effects of whole-body cryotherapy on serum mediators of inflammation
618 and serum muscle enzymes in athletes, *Journal of thermal biology* 34 (2)
619 (2009) 55–59.
- 620 [30] B. Dugué, E. Leppänen, Adaptation related to cytokines in man: Effects of
621 regular swimming in ice-cold water, *Clinical physiology (Oxford, England)* 20
622 (2) (2000) 114–121.

- 623 [31] M. Buemi, A. Allegra, C. Aloisi, F. Corica, A. Alonci, A. Ruello, G. Montalto,
624 N. Frisina, Cold Pressor Test Raises Serum Concentrations of ICAM-1,
625 VCAM-1, and E-Selectin in Normotensive and Hypertensive Patients,
626 Hypertension 30 (4) (1997) 845–847.
- 627 [32] G. Banfi, G. Melegati, A. Barassi, G.M. d’Eril, Effects of the whole-body
628 cryotherapy on NTproBNP, hsCRP and troponin I in athletes, Journal of
629 science and medicine in sport / Sports Medicine Australia 12 (6) (2009) 609–
630 610.
- 631 [33] P. Brancaccio, N. Maffulli, F.M. Limongelli, Creatine kinase monitoring in
632 sport medicine, British medical bulletin 81-82 (2007) 209–230.
- 633 [34] P. Brancaccio, G. Lippi, N. Maffulli, Biochemical markers of muscular
634 damage, Clinical chemistry and laboratory medicine 48 (6) (2010) 757–767.
- 635 [35] J.P. Hyatt, P.M. Clarkson, Creatine kinase release and clearance using MM
636 variants following repeated bouts of eccentric exercise, Medicine and science
637 in sports and exercise 30 (7) (1998) 1059–1065.
- 638 [36] M. van de Vyver, K.H. Myburgh, Cytokine and satellite cell responses to
639 muscle damage: Interpretation and possible confounding factors in human
640 studies, Journal of muscle research and cell motility 33 (3-4) (2012) 177–185.
- 641 [37] O. Neubauer, D. König, K.-H. Wagner, Recovery after an Ironman triathlon:
642 Sustained inflammatory responses and muscular stress, European journal of
643 applied physiology 104 (3) (2008) 417–426.
- 644 [38] E. Ziemann, R.A. Olek, S. Kujach, T. Grzywacz, J. Antosiewicz, T. Garsztko,
645 R. Laskowski, Five-day whole-body cryostimulation, blood inflammatory
646 markers, and performance in high-ranking professional tennis players,
647 Journal of athletic training 47 (6) (2012) 664–672.
- 648 [39] A. Wozniak, B. Wozniak, G. Drewa, C. Mila-Kierzenkowska, A. Rakowski,
649 The effect of whole-body cryostimulation on lysosomal enzyme activity in
650 kayakers during training, European journal of applied physiology 100 (2)
651 (2007) 137–142.
- 652 [40] G. Lombardi, P. Lanteri, S. Porcelli, C. Mauri, A. Colombini, D. Grasso, V.
653 Zani, F.G. Bonomi, G. Melegati, G. Banfi, Hematological profile and martial
654 status in rugby players during whole body cryostimulation, PloS one 8 (2)
655 (2013) e55803.
- 656 [41] L.D. Hayes, F.M. Grace, J.S. Baker, N. Sculthorpe, Exercise-induced
657 responses in salivary testosterone, cortisol, and their ratios in men: A meta-
658 analysis, Sports medicine (Auckland, N.Z.) 45 (5) (2015) 713–726.
- 659 [42] K.N. van Paridon, M.A. Timmis, C.M. Nevison, M. Bristow, The anticipatory
660 stress response to sport competition; a systematic review with meta-analysis
661 of cortisol reactivity, BMJ open sport & exercise medicine 3 (1) (2017)
662 e000261.
- 663 [43] P. Wahl, S. Mathes, K. Köhler, S. Achtzehn, W. Bloch, J. Mester, Acute
664 metabolic, hormonal, and psychological responses to different endurance
665 training protocols, Hormone and metabolic research 45 (11) (2013) 827–833.
- 666 [44] J.T. Costello, R.A. Rendell, M. Furber, H.C. Massey, M.J. Tipton, J.S. Young,
667 J. Corbett, Effects of acute or chronic heat exposure, exercise and

- 668 dehydration on plasma cortisol, IL-6 and CRP levels in trained males,
669 Cytokine (2018).
- 670 [45] D. Grasso, P. Lanteri, C. Di Bernardo, C. Mauri, S. Porcelli, A. Colombini, V.
671 Zani, F.G. Bonomi, G. Melegati, G. Banfi, G. Lombardi, Salivary steroid
672 hormone response to whole-body cryotherapy in elite rugby players, Journal
673 of biological regulators and homeostatic agents 28 (2) (2014) 291–300.
- 674 [46] A. Wozniak, C. Mila-Kierzenkowska, M. Szpinda, J. Chwalbinska-Moneta, B.
675 Augustynska, A. Jurecka, Whole-body cryostimulation and oxidative stress in
676 rowers: The preliminary results, Archives of medical science AMS 9 (2)
677 (2013) 303–308.
- 678 [47] K. Schaal, Y. Le Meur, J. Louis, J.-R. Filliard, P. Hellard, G. Casazza, C.
679 Hauswirth, Whole-Body Cryostimulation Limits Overreaching in Elite
680 Synchronized Swimmers, Medicine and science in sports and exercise 47 (7)
681 (2015) 1416–1425.
- 682